RESEARCH Open Access

Cytometric analysis of patients with COVID-19: what is changed in the second wave?

Giulia Scalia¹, Maddalena Raia¹, Monica Gelzo^{1,2}, Sara Cacciapuoti³, Annunziata De Rosa⁴, Biagio Pinchera³, Riccardo Scotto³, Agnese Giaccone³, Mauro Mormile³, Gabriella Fabbrocini³, Ivan Gentile³, Roberto Parrella⁴ and Giuseppe Castaldo^{1,2*}

Abstract

Background: Coronavirus disease 2019 (COVID-19) pandemic had a 1st wave in Europe from March to May 2020 and a 2nd wave since September 2020. We previously studied 35 hospitalized COVID-19 patients of the 1st wave demonstrating a cytokine storm and the exhaustion of most lymphocyte subpopulations. Herein, we describe the results obtained from COVID-19 patients of the 2nd wave.

Methods: We analyzed interleukin (IL)-6 by human-specific enzyme-linked immunosorbent assay and a large set of lymphocyte subpopulations by flow cytometry in 274 COVID-19 patients hospitalized from September 2020 to May 2021.

Results: Patients of 2nd wave compared with those of 1st wave showed lower serum IL-6 levels and a higher number of B and most T lymphocyte subpopulations in advanced stages, in relation with the age and the gender. On the other hand, we observed in 2nd wave patients: (i) a reduction of most lymphocyte subpopulations at mild and moderate stages; (ii) a reduction of natural killer cells and T regulatory cells together with a higher number of activated T helper (TH) 17 lymphocytes in all stages, which were mainly related to steroid and azithromycin therapies before hospitalization.

Conclusions: COVID-19 had a less severe impact in patients of the 2nd wave in advanced stages, while the impact appeared more severe in patients of mild and moderate stages, as compared with 1st wave patients. This finding suggests that in COVID-19 patients with milder expression at diagnosis, steroid and azithromycin therapies appear to worsen the immune response against the virus. Furthermore, the cytometric profile may help to drive targeted therapies by monoclonal antibodies to modulate specific IL/lymphocyte inhibition or activation in COVID-19 patients.

Keywords: Cytometry, Interleukin-6, Lymphocytes, Targeted therapy, COVID-19

Introduction

Coronavirus disease 2019 (COVID-19) may appear with a widely variable clinical expression, from asymptomatic or mild [1] to severe forms [2] with acute respiratory distress syndrome (ARDS) and multi-organ failure in less than 10% of patients. The COVID-19 pandemic had a 1st wave in Italy from March to May 2020. To cast light on immunological and cytological response, we

Full list of author information is available at the end of the article



^{*}Correspondence: giuseppe.castaldo@unina.it

¹ CEINGE-Biotecnologie Avanzate, via Gaetano Salvatore 486, 80145 Naples, Italy

Scalia et al. J Transl Med (2021) 19:403 Page 2 of 9

performed a preliminary study on 35 patients hospitalized for COVID-19 demonstrating a reduction of lymphocytes including both B and T populations. Among T lymphocytes, we observed a reduction of helper, suppressor and regulatory subpopulations, in relation with the disease severity [3]. Serum IL-6 was increased in most patients with a significant association with the disease severity. Thus, the analysis demonstrated the picture of lymphocyte exhaustion induced by the cytokine storm. The molecular mechanisms that cause these complications only in a small subset of COVID-19 patients are still under study [4]. In fact, even if most lymphocyte subpopulations are depleted, some T cell subgroups, involved in the synthesis of cytokines, are strongly activated in few severe COVID-19 patients [5, 6], creating a vicious circle between the cytokine storm and lymphocyte exhaustion [7]. Thus, the analysis of lymphocyte subpopulations in COVID-19 patients could contribute to predict the outcome and to define personalized therapies targeting different cell or cytokine pathways on the basis of cytometric analysis [3, 8, 9].

After a lockdown during summer 2020, the pandemic in Italy had a 2nd wave since September 2020. The two pandemic waves had several differences [10-12]. In fact, during the 1st wave most patients were diagnosed after the onset of symptoms by molecular analysis on nasopharyngeal swab that often required 2 or 3 days, and the patients were hospitalized soon, after the result. While, most patients of the 2nd wave were diagnosed when they were still asymptomatic because they had been traced following a contact with a COVID-19 patient. The result of the nasopharyngeal test was obtained more rapidly and some patients began to be treated with different combinations of steroids, azithromycin, and heparin several days before hospitalization [13-15]. During the 2nd wave, we studied 274 novel COVID-19 patients recovered in our specialized hospitals with the same protocol of the first study [3]. We now describe the results, in comparison with those obtained in patients from the 1st wave.

Methods

Patients

All 274 consecutive adult patients (mean age: 61.2 years, range: 17–91 years; 135 females, 49.0%) with a diagnosis of COVID-19 admitted at one of our hospitals from September 2020 to May 2021 (2nd wave of COVID-19 pandemic) were enrolled. The lone exclusion criterion was the refusal or the impossibility to sign the informed consent. None of the patients admitted to our Institutions during the period of our study was excluded. The diagnosis of COVID infection was confirmed by molecular analysis on nasopharingeal

swab [16]. All the enrolled patients were classified on the basis of the seven ordinal scale made by the World Health Organization (WHO)-Research and Development Blueprint expert group and used in previous influenza studies [17, 18]. Furthermore, for each patient we recorded the assumption of drugs (particularly steroids, azithromycin or both drugs) before hospitalization. As shown in Table 1, 161/274 (59.0%) patients were treated with one or both the drugs before hospitalization.

Lymphocyte subpopulations and serum IL-6 analyses

Whole blood samples were collected at admission in tubes containing EDTA and then immediately analyzed by flow cytometry. Serum samples were separated from blood cells after the collection in tubes without anticoagulant and stored at $-80\,^{\circ}\text{C}$ until IL-6 analysis [3]. Immunophenotyping analysis was performed by multicolour flow cytometry [3]. Serum IL-6 levels were analyzed using human-specific enzyme-linked immunosorbent assay (ELISA) Max^{TM} Set Deluxe kits (BioLegend, Inc., San Diego, USA), in accordance with the manufacturer's instructions.

Statistical analysis

The data obtained in the 274 patients of the 2nd wave were compared to those collected in patients of the 1st wave [3]. Continuous data were reported as median and interquartile range (IQR). Comparisons between two groups were evaluated by Mann-Whitney U test. Statistical differences between three groups were assessed by Kruskal-Wallis test and Mann-Whitney U test as posthoc test. Categorical data were reported as frequency and percentage. The chi-square test was used to compare the frequency of categorical variables between the groups. Linear regression analysis was used to assess the effects of age, gender and therapies on lymphocyte subpopulations. Statistical analysis was performed by SPSS (version 26, IBM SPSS Statistics). Graphics have been performed by KaleidaGraph software (version 4.5.4, Synergy, Reading, PA, USA). P values < 0.05 were considered as significant.

 Table 1
 Steroids
 and/or
 azithromycin
 therapies
 before

 hospitalization in 2nd wave patients

Subgroups	Not treated	Only steroids	Only azithromycin	Steroids and azithromycin
WHO 3	53 (81)	5 (8)	2 (3)	5 (8)
WHO 4	44 (31)	21 (15)	6 (4)	70 (50)
WHO 5-7	16 (24)	13 (19)	5 (7)	34 (50)

N (%)

Scalia et al. J Transl Med (2021) 19:403 Page 3 of 9

Results

As shown in Table 2, the WHO stage distribution of COVID-19 patients of the two waves was not significantly different. In contrast, the age of patients of the 2nd wave was significantly lower, even if in both waves the age of patients increased (significantly in patients of the 2nd wave) with parallel to the WHO stage. Furthermore, although among patients of the 2nd wave we observed a significantly lower number of males as compared to the 1st wave, the percentage of males increased with the progression of the WHO stage.

In addition, the data of serum IL-6 and lymphocyte subpopulations in 274 COVID patients of the 2nd wave in comparison with the 35 COVID patients of the 1st wave [3] are reported in Table 2. We describe the comparison of the data among the WHO subgroups of patients of the same wave as well as the comparison between the two waves patients bearing to the same WHO subgroup. Comparing the patients of the two waves, in patients of the 2nd wave we observed that:

- (i) Serum IL-6 was significantly lower in each WHO subgroup, with no differences among WHO stages. While, in patients of the 1st wave the levels of the marker showed an increasing trend with the stage [3].
- (ii) Total lymphocytes (Fig. 1A) were lower in patients of the WHO stage 3 and 4, while they resulted higher (always not significantly) in patients of the WHO stages 5–7. Furthermore, the trend of reduction with the stage was less pronounced than in patients of the 1st wave.
- (iii) T lymphocytes (Fig. 1B) were not different in patients of the WHO stages 3 and 4, while they resulted higher (although not significantly) in patients of stages 5-7. Furthermore, the trend of reduction with the stage was less pronounced than in patients of the 1st wave. T helper and T suppressor lymphocytes were lower in patients of the WHO stage 3, while they resulted not different in patients of the WHO stage 4 and higher (although not significantly) in patients of the WHO stage 5-7. Furthermore, for both the parameters we observed the same significant reduction with the progression of the WHO stage observed in patients of 1st wave [3]. (iv) B lymphocytes (Fig. 1C) were lower in patients of the WHO 3 stage, while they resulted higher in patients of the WHO stage 4 (significantly) and of the stage 5-7. Furthermore, B lymphocytes did not show the trend of reduction with the severity that

we observed in patients of the 1st wave [3]. Naïve lymphocytes were slightly lower in patients of the WHO stage 3 and higher in patients of the WHO stages 4 and 5–7 (although not significantly). Furthermore, the trend of reduction with the stage was less pronounced than in patients of the 1st wave.

- (v) NK lymphocytes (Fig. 2A) resulted lower in patients from all WHO stages (significantly for the WHO 4 subgroup) with no differences between subgroups, while in patients of the 1st wave they gradually reduced with the progression of the WHO stage (Fig. 2A). T regulatory lymphocytes (Fig. 2B) were lower in each WHO stage (significantly in subgroups 3 and 4), with the same trend of significant reduction with the increase of the WHO stage.
- (vi) Total activated (Fig. 3A), as well as activated T lymphocytes, were lower in patients of the WHO stage 3, significantly higher (total activated) or equal (activated T) in WHO stage 4, and both increased (although not significantly) in patients of the WHO stage 5–7.
- (vii) Both TH1 and TH17 lymphocytes were lower in WHO stage 3, equal in WHO stage 4 and higher (although not significantly) in WHO stage 5–7. Both the subpopulations had a trend of reduction with the stage (significant for TH1) in both 1st wave and 2nd wave.
- (viii) Activated TH1 (Fig. 3B) lymphocytes were lower (although not significantly) in patients of all WHO stages, while activated TH17 lymphocytes (Fig. 3C) resulted higher (although not significantly) in all WHO stages and did not show any trend with the increase of the WHO stage. Furthermore, we evaluated the ratio between activated TH1 and TH17 lymphocytes in patients of both the waves (Fig. 3D).

Finally, we performed the linear regression analysis to evaluate the effect of age, gender and pre-recovery treatments on the values of lymphocyte populations (Table 3). Both the age and the male gender were negatively related to the number of most lymphocyte subpopulations (i.e., total, T, T helper, T suppressor, naïve, T regulatory and TH1 lymphocytes). Furthermore, we observed a negative correlation between the assumption of steroids and the number of T suppressor, NK and T regulatory lymphocytes, and a negative correlation between the assumption of azithromycin and T, T regulatory and TH1 lymphocytes number.

Scalia et al. J Transl Med (2021) 19:403 Page 4 of 9

Table 2 Comparison of demographics, serum IL-6 (pg/mL) and circulating lymphocytes (N/mmc) in COVID-19 patients of 1st wave and patients of 2nd wave at admission with different severity according to worst WHO stage for each patient

	Wave	All	WHO 3	WHO 4	WHO 5-7	Kruskal- Wallis
N (%)	1st	35	7 (20)	20 (57)	8 (23)	
	2nd	274	65 (24)	141 (51)	68 (25)	_
	1st vs 2nd	_	n.s	n.s	n.s	
Age, yrs	1st	62 (50-73)	60 (39–62)	64 (51–73)	75 (57–79)	n.s
	2nd	55 (37–66)	34 (29–48)	59 (41–68) a	57 (48–73) ^b	< 0.0001
	1st vs 2nd	0.023	n.s	n.s	n.s	
Males, N (%)	1st	27 (77)	4 (57)	16 (80)	7 (88)	_
	2nd	139 (51)	15 (23)	77 (55)	47 (69) ^b	_
	1st vs 2nd	0.003	n.s	0.031	n.s	
IL-6	1st	171 (90–397)	130 (90-223)	197 (86-375)	292 (53-769) ^b	0.021
0–4.5	2nd	28 (23–41)	28 (23-41)	28 (21–41)	31 (25–46)	n.s
	1st vs 2nd	< 0.0001	0.0007	< 0.0001	0.004	
Total	1st	1116 (539–1387)	1595 (588–1924)	1153 (653–1333)	450 (267-1033)	n.s
1500–3000	2nd	1058 (709–1683)	1289 (874–1860)	1058 (736–1577)	770 (518–1735)	n.s
	1st <i>vs</i> 2nd	n.s	n.s	n.s	n.s	
Т	1st	793 (395–1027)	959 (306–1673)	801 (485–1015) ^a	309 (221–776) ^b	0.041
605–2460	2nd	801 (465–1245)	996 (600–1477)	780 (495–1187)	564 (336–1203)	n.s
	1st <i>vs</i> 2nd	n.s	n.s	n.s	n.s	
T helper	1st	482 (225–692)	692 (223–803)	527 (268–616)	215 (162–421) ^b	0.050
493–1666	2nd	461 (283–797)	539 (321–873)	461 (316–794)	364 (173–702) ^{b, c}	0.040
	1st vs 2nd	n.s	n.s	n.s	n.s	
T suppressor	1st	201 (109–357)	304 (56–482)	197 (123–357)	88 (30–310)	n.s
229–1112	2nd	249 (146–395)	287 (204–436)	236 (148–361)	198 (102–376) ^b	0.018
	1st vs 2nd	n.s	n.s	n.s	n.s	
В	1st	76 (40–154)	154 (47–241)	67 (37–119)	68 (38–106)	n.s
72–520	2nd	141 (67–244)	106 (64–225)	160 (77–247)	121 (47–272)	n.s
	1st vs 2nd	0.004	n.s	0.009	n.s	
Naïve	1st	463 (287–915)	861 (412–1231)	468 (337–811)	252 (191–788)	n.s
126–1121	2nd	648 (431–1071)	759 (510–1180)	688 (440–1008)	482 (328–1077)	n.s
	1st vs 2nd	n.s	n.s	n.s	n.s	
NK	1st	134 (84–239)	153 (117–239)	148 (99–266)	88 (71–128)	n.s
73–654	2nd	81 (48–132)	93 (51–157)	77 (47–125)	80 (48–130)	n.s
	1st vs 2nd	0.004	n.s	0.002	n.s	
T regulatory	1st	17.6 (7.0–27.3)	24.5 (17.6–35.1)	18.9 (8.4–31.4)	9.1 (3.2–17.1) ^b	0.03
7-52	2nd	9.1 (4.8–15.5)	11.0 (7.3–18.3)	9.3 (4.7–15.4) ^a	7.1 (2.96–12.7) ^b	< 0.0001
	1st vs 2nd	0.002	0.024	0.008	n.s	
Total activated	1st	137 (88–234)	234 (100–478)	137 (113–209)	87 (53–223)	n.s
86-799	2nd	193 (104–311)	171 (99–309)	206 (127–311)	138 (85–324)	n.s
	1st vs 2nd	n.s	n.s	0.034	n.s	
Activated T	1st	30.1 (11.8–56.9)	37.1 (6.1–98.9)	31.3 (19.6–54.5)	10.4 (7.2–61.9)	n.s
14–411	2nd	25.0 (12.6–45.8)	24.7 (12.0–46.6)	31.0 (15.6–48.6)	19.9 (11.0–35.1)	n.s
•	1st vs 2nd	n.s	n.s	n.s	n.s	
TH1	1st	93 (49–195)	180 (56–249)	89 (51–192)	44 (19–107) ^b	0.050
37–220	2nd	79 (42–147)	99 (65–178)	80 (44–139)	55 (27–117) ^{b, c}	0.016
-:	1st <i>vs</i> 2nd	n.s	n.s	n.s	n.s	
Activated TH1	1st vs 211d	6.3 (2.7–12.2)	6.4 (2.4–15.7)	6.2 (2.7–12.4)	5.3 (1.3–8.9)	n.s
0–20	2nd	3.6 (1.7–7.6)	4.0 (1.6–6.9)	4.4 (2.0–10.3)	3.1 (1.6–6.1)	n.s
0 20	2110	5.0 (1.7 7.0)	7.0 (1.0 '0.2)	T.T (2.0 10.3)	J.1 (1.0 ·0.1)	11.3

Scalia et al. J Transl Med (2021) 19:403 Page 5 of 9

Table 2 (continued)

	Wave	All	WHO 3	WHO 4	WHO 5-7	Kruskal- Wallis
TH17	1st	44.2 (22.3–65.1)	55.4 (22.3–66.0)	50.4 (27.6–66.1)	22.8 (12.0–42.0)	n.s
3.76–60.0	2nd	45.2 (24.1–76.6)	46.3 (19.6–70.6)	50.3 (26.1–80.8)	38.0 (20.6–76.9)	n.s
	1st vs 2nd	n.s	n.s	n.s	n.s	
Activated TH17	1st	0.57 (0.23-0.80)	0.69 (0.45-0.80)	0.55 (0.15-0.89)	0.40 (0.24-0.73)	n.s
0-1.3	2nd	0.63 (0.00-1.44)	0.53 (0.00-1.29)	0.71 (0.11-1.66)	0.58 (0.00-1.76)	n.s
	1st vs 2nd	n.s	n.s	n.s	n.s	
Activated TH1/ TH17	1st	12 (5.8–38)	10 (6.0–29)	12 (5.2–98)	11 (3.9–31)	n.s
	2nd	7.0 (3.0-27)	8.0 (2.2-92)	7.3 (3.5–35)	6.5 (3.0–16)	n.s
	1st vs 2nd	n.s	n.s	n.s	n.s	

Median and interquartile range. For each parameter we report the reference range

n.s. not significant

Significant values are reported in bold

Discussion

Patients of the 2nd wave were significantly younger as compared with patients of the 1st wave, which have been described in our preliminary study [3], even if, in both the waves, the age of patients significantly increased with parallel to the WHO stage. Furthermore, among patients of the 2nd wave we observed a significantly higher percentage of females, although in advanced stages from both the waves the percentage of males was higher, indicating that age and male gender are risk factors for a more severe disease. In fact, the 10/12 (83.0%) patients of the 2nd wave, that died during hospitalization, were males (data not shown), in disagreement with a large Italian study that reported a higher percentage of females among patients died for COVID-19 during the 2nd wave [11].

Furthermore, in COVID-19 patients of the 2nd wave we observed a higher number of total, T, helper, suppressor, naïve and B lymphocytes in patients in advanced WHO stages (5 to 7), while in the other patients all (stage 3) or some (stage 4) of the above-mentioned subpopulations were lower in comparison to patients of the 1st wave of the corresponding WHO stage. These findings indicate that the 2nd wave was less severe only for patients in advanced stage, in disagreement with previous studies that reported a global less severe clinical impact of the 2nd wave of COVID-19. The younger age and the higher number of females that we observed in patients of the 2nd wave in comparison with patients of 1st wave, may depend on the improvement of organizational and diagnostic strategies that allow the early hospitalization of COVID-19 patients. The improvement might also depend on the steroid therapy performed before hospitalization (about 76% of patients of stages 5-7), while all patients of the 1st wave were hospitalized before starting therapies. Steroids could contribute to reduce the cytokine storm inhibiting the production of IL-6 [19, 20] thus rendering less pronounced the lymphocyte exhaustion [9], even if our regression analysis excluded the impact of steroid therapy on IL-6 serum levels.

While, the less encouraging results that we obtained in patients of WHO stages 3 (about 35% of which were treated before hospitalization) and 4 (about 70% treated before hospitalization) may depend on the prevalent inhibitory effect on lymphocyte proliferation that steroids and azithromycin exerted in patients with a less severe COVID-19 disease. In fact, the current literature reports that steroid therapy should be avoided in COVID-19 patients not progressed to a stage that requires oxygen support (like all our patients of WHO stages 3 and 4 before hospitalization), because the immunosuppressive effects of these drugs might hamper antiviral response [13, 19]. While, the pharmacological role of azithromycin is still questioned [21], given the weak evidences of its antiviral effect in COVID-19 patients [14]. In addition, all patients of the 2nd wave had a reduction of NK cells that have a relevant role in clearing virus-infected cells [8, 22] and of T regulatory lymphocytes. This reduction may depend on a higher number of T regulatory lymphocytes infected by the virus and thus removed [5], or on a higher number of cells migrated in lung [6], but may also depend on steroids assumed before hospitalization that inhibit the NK response [22] and reduce the T regulatory production [23]. The relevant reduction of T regulatory lymphocytes in COVID-19 patients was observed also in other studies [5]. Considering the immunomodulatory role of these cells in keeping in check inflammation, various therapeutic approaches have been proposed

 $^{^{}a}$ p < 0.01, WHO 4 versus WHO 3; b p < 0.01, WHO 5–7 versus WHO 3; c p < 0.01, WHO 5–7 versus WHO 4

Scalia et al. J Transl Med (2021) 19:403 Page 6 of 9

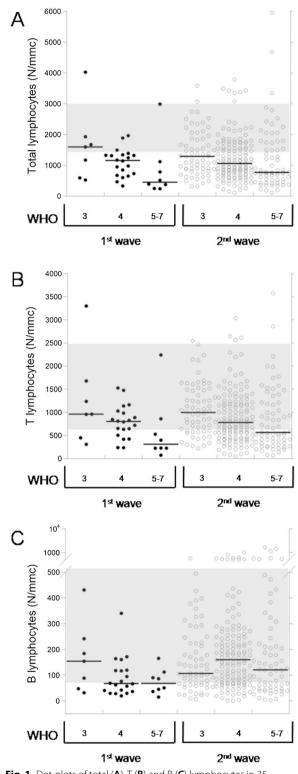


Fig. 1 Dot-plots of total **(A)**, T **(B)** and B **(C)** lymphocytes in 35 patients of COVID-19 1st wave and 274 patients of the 2nd wave at hospital admission. Gray areas indicate the reference ranges. Black lines indicate median values

to stimulate the production of such cells [5]. Cytometric analysis may help to specifically address these therapies to patients with a lower number of T regulatory lymphocytes.

Furthermore, we observed a lower number of activated TH1 lymphocyte and an increase of activated TH17 cells particularly in patients with advanced WHO stages, despite the reduction of neutrophils [24] that should mediate TH17 promotion [25]. In addition, these results may be influenced by the therapies that our patients performed before hospitalization. In fact, it is known that both steroids [26] and azithromycin modify the TH axis suppressing activated TH1 and TH1 lymphocytes [27] and related chemokines like IFN-gamma impairing the antiviral response [28-30]. Thus, the TH1/TH17 balance is addressed toward a pro-inflammatory condition. Various studies suggested the inhibitory targeting of TH17, IL-17 or IL-17RA and the use of TH1 activators as potential therapeutic approaches for COVID-19 [31-35]. IL-17 inhibition has already been adopted as a common and successful strategy to reduce the injury associated with inflammatory autoimmune diseases including psoriasis and psoriatic arthritis. Dysregulation of TH17 cells and production of IL-17 in the skin, synovial space and endothelium promote the production of downstream pro-inflammatory molecules such as IL-1β, TNF and IL-6. Recruited neutrophils then produce IL-6 and reactive oxygen species, leading to characteristic skin lesions and joint destruction. In ARDS and in the acute lung injury, such us in psoriasis, there is a disruption of the balance of pro-inflammatory and anti-inflammatory cytokines [34]. Three commercially available anti IL-17 drugs exist: secukinumab (human monoclonal antibody to IL-17) [36], ixekizumab (humanized monoclonal antibody to IL-17) and brodalumab (human monoclonal antibody to the IL-17 receptor). Both secukinumab and ixekizumab are approved for psoriasis, psoriatic arthritis and ankylosing spondylitis; brodalumab is approved for the treatment of psoriasis alone [37]. By targeting IL-17, which operates 'upstream' of both IL-1 and IL-6 and results in a reduction of neutrophil recruitment, several factors known to play major roles in ARDS would be inhibited [34].

Our study demonstrates a strong variability of TH1/TH17 response among COVID-19 patients also within the same WHO stage. Thus, cytometric analysis may help to treat each patient with the most appropriate targeted approach by a monoclonal antibody treatment against the altered immune pathway, such as tocilizumab for IL-6 pathway [38] or secukinumab for IL-17 pathway [3, 33] alterations.

Scalia et al. J Transl Med (2021) 19:403 Page 7 of 9

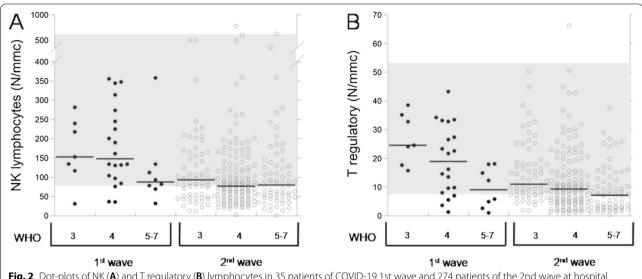


Fig. 2 Dot-plots of NK (A) and T regulatory (B) lymphocytes in 35 patients of COVID-19 1st wave and 274 patients of the 2nd wave at hospital admission. Gray areas indicate the reference ranges. Black lines indicate median values

To conclude: our study indicates that in COVID-19 patients of the 2nd wave, the younger age, the prevalence of the females and steroid therapy before hospitalization have a global positive effect in patients with advanced WHO stages. While, the assumption of steroids and azithromycin at diagnosis has a negative impact on cytometric parameters in less severe patients worsening the immune response against the virus. A limitation of this

study is represented by the low number of patients and the comparison between obviously non-homogeneous groups. Further studies are necessary to better define the therapies to be used in COVID-19 patients with different severity and to reveal prognostic biomarker that early predict the severity of the disease. In this context, cytometric analysis may contribute to select targeted therapies by monoclonal antibodies for each patient.

Table 3 Linear regression analysis in 2nd wave COVID-19 patients

	Age		Gender (male)		Steroids*		Azithromycin*	
	Slope	P value	Slope	P value	Slope	P value	Slope	P value
IL-6	- 0.180	0.064	- 0.078	0.255	- 0.054	0.324	- 0.055	0.322
Total	- 0.206	< 0.0001	- 0.133	0.015	- 0.097	0.058	- 0.095	0.061
Т	- 0.249	< 0.0001	- 0.170	0.003	- 0.096	0.059	- 0.113	0.033
T helper	- 0.212	< 0.0001	- 0.155	0.006	- 0.042	0.250	- 0.097	0.058
T suppressor	- 0.211	< 0.0001	- 0.115	0.030	- 0.113	0.033	- 0.068	0.134
В	- 0.089	0.075	- 0.045	0.232	- 0.013	0.414	- 0.005	0.468
Naïve	- 0.225	< 0.0001	- 0.121	0.024	- 0.100	0.052	- 0.084	0.086
NK	- 0.034	0.290	0.032	0.301	- 0.111	0.035	- 0.081	0.095
T regulatory	- 0.294	< 0.0001	- 0.211	< 0.0001	- 0.192	0.001	- 0.153	0.006
Total activated	- 0.094	0.064	- 0.058	0.175	- 0.028	0.323	- 0.012	0.422
Activated T	0.000	0.498	- 0.049	0.212	0.039	0.264	0.027	0.328
TH1	- 0.235	< 0.0001	- 0.224	< 0.0001	- 0.083	0.089	- 0.119	0.027
Activated TH1	0.040	0.259	- 0.027	0.332	0.043	0.245	- 0.016	0.395
TH17	- 0.078	0.102	0.052	0.201	- 0.006	0.463	- 0.085	0.084
Activated TH17	0.050	0.211	0.040	0.261	0.031	0.307	- 0.004	0.477
Activated TH1/TH 17	- 0.067	0.138	- 0.083	0.090	- 0.024	0.346	0.017	0.390

^{*} Among the 274 COVID-19 patients of the 2nd wave, 161 have been treated with steroids and/or azithromycin before hospitalization Significant values are reported in bold

Scalia et al. J Transl Med (2021) 19:403 Page 8 of 9

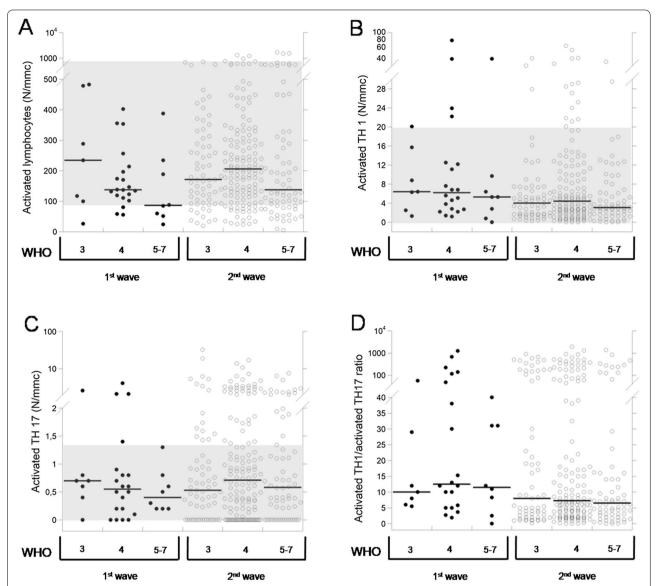


Fig. 3 Dot-plots of total activated (A), activated TH1 (B), activated TH17 (C) lymphocytes and activated TH1/activated TH17 ratio in 35 patients of COVID-19 1st wave and 274 patients of the 2nd wave at hospital admission. Gray areas indicate the reference ranges. Black lines indicate median values

Acknowledgements

We thank the staff of CEINGE-Biotecnologie Avanzate that hosts the scientific activity of the project.

Authors' contributions

GF, IG, RP, and GC designed the study. SC, ADR, BP, RS, and AG recruited patients and provided samples and clinical data. GS, MR, and MG performed IL-6 and cytometric analysis. MG and GC performed statistical analysis. MM, GF, IG, RP, and GC wrote the manuscript. All authors participated in preparation of the manuscript and gave final approval for publication. All authors have read and approved the final manuscript.

Funding

This research was supported by Regione Campania (CEINGE-TASK-force COVID-19, code D64I200003800).

Availability of data and materials

All data used during the study are available from the corresponding author by request.

Declarations

Ethics approval and consent to participate

Ethical approval for the study was obtained from the Ethical Committee of the University Federico II of Naples (protocol code 138/20, April 14, 2020). All experiments were performed in accordance with relevant guidelines and regulations. The study has been performed in accordance with the Declaration of Helsinki. Informed consent was obtained from all participants.

Competing interests

The authors declare that they have no competing interests.

Scalia et al. J Transl Med (2021) 19:403 Page 9 of 9

Author details

¹CEINGE-Biotecnologie Avanzate, via Gaetano Salvatore 486, 80145 Naples, Italy. ²Dipartimento di Medicina Molecolare e Biotecnologie Mediche, Università di Napoli Federico II, Naples, Italy. ³Dipartimento di Medicina Clinica e Chirurgia, Università di Napoli Federico II, Naples, Italy. ⁴Dipartimento di Malattie Infettive e Emergenze Infettive, Divisione di malattie infettive respiratorie, Ospedale Cotugno, AORN dei Colli, Naples, Italy.

Received: 19 July 2021 Accepted: 7 September 2021 Published online: 23 September 2021

References

- Gandhi RT, Lynch JB, Del Rio C. Mild or moderate COVID-19. N Engl J Med. 2020;383(18):1757–66.
- Berlin DA, Gulick RM, Martinez FJ. Severe covid-19. N Engl J Med. 2020;383(25):2451–60.
- Cacciapuoti S, De Rosa A, Gelzo M, Megna M, Raia M, Pinchera B, et al. Immunocytometric analysis of COVID patients: a contribution to personalized therapy? Life Sci. 2020;261:118355.
- Berthelot JM, Liote F, Maugers Y, Sibilia J. Lymphocyte changes in severe COVID-19: delayed over-activation of STING? Front Immunol. 2020:11:607069.
- Strephen-Victor E, Das M, Karnam A, Pitard B, Gautier JF, Bayry J. Potential of regulatory T-cell-based therapies in the management of severe COVID-19. Eur Resp J. 2020;56(3):2002182.
- Delshad M, Tavakolinia N, Pourbagheri-Sigaroodi A, Safaroghil-Azar A, Bagheri N, Bashash D. The contributory role of lymphocyte subsets, pathophysiology of lymphopenia and its implication as prognostic and therapeutic opportunity in COVID-19. Intern Immunopharmacol. 2021. https://doi.org/10.1016/j.intimp.2021.107586.
- Zenarruzabeitia O, Astarloa-Pando G, Terrén I, Orrantia A, Perez-Garay R, Seijas-Bertolaza I, et al. T cell activation, highly armed cytotoxic cells and a shift in monocytes CD300 receptors expression is characteristics of patients with severe COVID-19. Front Immunol. 2021;12:655934.
- Bozzano F, Dentone C, Perrone C, Di Biagio A, Fenoglio D, Parodi A, et al. Extensive activation, tissue trafficking, turnover and functional impairment of NK cells in COVID-19 patients at disease onset associates with subsequent disease severity. PLoS Pathog. 2021;17(4):e1009448.
- De Candia P, Prattichizzo F, Garavelli S, Matarese G. T cells: warriors of SARS-CoV-2 infection. Trends Immunol. 2021;42(1):18–30.
- De Natale G, De Natale L, Troise C, Marchitelli V, Coviello A, Holmberg KG, et al. The evolution of Covid-19 in Italy after the spring of 2020: an unpredicted summer respite followed by a second wave. Int J Environ Res Public Health. 2020;17(23):8708.
- Palmieri L, Palmer K, Lo Noce C, Meli P, Giuliano M, Floridia M, Italian National Institute of Health COVID-19 Mortality Group, et al. Differences in the clinical characteristics of COVID-19 patients who died in hospital during different phases of the pandemic: national data from Italy. Aging Clin Exp Res. 2021;33(1):193–9.
- Borghesi A, Golemi S, Carapella N, Zigliani A, Farina D, Maroldi R. Lombardy, Northern Italy: COVID-19 second wave less severe and deadly than the first? A preliminary investigation. Infect Dis. 2021;53(5):370–5.
- 13. The RECOVERY collaborative group. Dexamethasone in hospitalized patients with COVID-19. N Engl J Med. 2021;384(8):693–704.
- Sultana J, Cutroneo PM, Crisafulli S, Puglisi G, Caramori G, Trifirò G.
 Azithromycin in COVID-19 patients: pharmacological mechanism clinical evidence and prescribing guidelines. Drug Saf. 2020;43(8):691–8.
- Capoluongo ED, Amato F, Castaldo G. The friendly use of chloroquine in the COVID-19 disease: a warning for the G6PD-deficient males and for the unaware carriers of pathogenic alterations of the G6PD gene. Clin Chem Lab Med. 2020;58(7):1162–4.
- Zollo M, Ferrucci V, Izzo B, Quarantelli F, Domenico CD, Comegna M, et al. SARS-CoV-2 subgenomic N (sgN) transcripts in oro-nasopharingeal swabs correlate with the highest viral load, as evaluated by five different molecular methods. Diagnostics. 2021;11(2):288.
- 17. von Cube M, Grodd M, Wolkewitz M, Hazard D, Wengenmayer T, Canet E, et al. Harmonizing heterogeneous endpoints in coronavirus disease 2019 trials without loss of information. Crit Care Med. 2021;49(1):e11–9.

- WHO Working Group on the Clinical Characterization and Management of COVID-19 infection. A minimal common outcome measure set for COVID-19 clinical research. Lancet Infect Dis. 2020;20(8):e192–7.
- Cain DW, Cidlowski JA. After 62 years of regulating immunity, dexamethasone meets COVID-19. Nat Rev Immunol. 2020;20(10):587–8.
- Xiang Z, Liu J, Shi D, Chen W, Li J, Yan R, et al. Glucocorticoids improve severe or critical COVID-19 by activating ACE2 and reducing IL-6 levels. Int J Biol Sci. 2020;16(13):2382–91.
- Zimmermann P, Ziesenitz VC, Curtis N, Ritz N. The immunomodulatory effects of macrolides—a systematic review of the underlying mechanisms. Front Immunol. 2018;9:302.
- 22. Osman M, Faridi RM, Sigl W, Shabani-Rad MT, Dharmani-Khan P, Parker A, et al. Impaired natural killer cell counts and cytolytic activity in patients with severe COVID-19. Blood Adv. 2020;4(20):5035–9.
- Siomkajlo M, Mizera L, Szymczak D, Kolackov K, Grzegrxolka J, Bolanowski M, et al. Effect of systemic steroid therapy in Graves' orbitopathy on regulatory T cells and Th17/Treg ratio. J Endocrinol Invest. 2021. https:// doi.org/10.1007/s40618-021-01565-w.
- Gelzo M, Cacciapuoti S, Pinchera B, De Rosa A, Cernera G, Scialò F, et al. Prognostic role of neutrophil to lymphocyte ratio in COVID-19 patients: still valid in patients that had started therapy? Front Pub Health. 2021. https://doi.org/10.3389/fpubh.2021.664108.
- Parackova Z, Bloomfield M, Klocperk A, Sediva A. Neutrophils mediate Th17 promotion in COVID-19 patients. J Leukoc Biol. 2021;109(1):73–6.
- 26. Elenkov IJ. Glucocorticoids and the Th1/Th2 balance. Ann NY Acad Sci. 2004;1024:138–46.
- Kuo CH, Lee MS, Kuo HF, Lin YC, Hung CH. Azithromycin suppresses Th1and Th2-related chemokines IP-10/MDC in human monocytic cell line. J Microbiol Immunol Infect. 2019;52(6):872–9.
- Ansari AW, Sharif-Askari FS, Jayakumar MN, Mohammed AK, Sharif-Askari NS, Venkatachalam T, et al. Azithromycin differentially alters TCR-activated helper T cell subset phenotype and effector function. Front Immunol. 2020:11:556579
- Diao B, Wang C, Tan Y, Chen X, Liu Y, Ning L, et al. Reduction and functional exhaustion of T cells in patients with coronavirus disease 2019 (COVID-19). Front Immunol. 2020;11:827.
- Zheng M, Gao Y, Wang G, Song G, Liu S, Sun D, et al. Functional exhaustion of antiviral lymphocytes in COVID-19 patients. Cell Mol Immunol. 2020;17(5):533–5.
- 31. Toor SM, Saleh R, Sasidharan Nair V, Taha RZ, Elkord E. T-cell responses and therapies against SARS-CoV-2 infection. Immunology. 2020;162(1):30–43.
- Orlov M, Wander PL, Morrell ED, Mikacenic C, Wurfel MM. A case for targeting Th17 cells and IL-17A in SARS-CoV-2 infections. J Immunol. 2020;205(4):892–8.
- Bulat V, Situm M, Azdajic MD, Likic R. Potential role of IL-17 blocking agents in the treatment of severe COVID-19. Br J Clin Pharmacol. 2021;87(3):1578–81.
- 34. Pacha O, Sallman MA, Evans SE. COVID-19: a case for inhibiting IL.17? Nat Rev Immunol. 2020;20(6):345–6.
- Scavone C, Mascolo A, Rafaniello C, Sportiello L, Trama U, Zoccoli A, et al. Therapeutic strategies to fight COVID-19: which is the status artis? Br J Pharmacol. 2021. https://doi.org/10.1111/bph.15452.
- Costanzo A, Russo F, Galluzzo M, Stingeni L, Scuderi R, Zichichi L, et al. Secukinumab exhibits sustained and stable response in patients with moderate-to-severe psoriasis: results from the SUPREME study. Acta Derm Venereol. 2021. https://doi.org/10.2340/00015555-3816.
- Conti A, Peccerillo F, Amerio P, Balato A, Bardazzi F, Bianchi L, et al. Efficacy and safety of switching to ixekizumab in secukinumab nonresponder patients with psoriasis: results from a multicentre experience. Br J Dermatol. 2019;180(6):1547–8.
- Xu X, Han M, Li T, Sun W, Wang D, Fu B, et al. Effective treatment of severe COVID-19 patients with tocilizumab. Proc Natl Acad Sci USA. 2020;117(20):10970–5.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.