

LETTER TO THE EDITOR

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Biomarkers of COVID-19 severity may not serve patients with polycystic ovary syndrome

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To the Editor:

In a cohort of patients with differing severity of COVID-19 disease, including non-survivors, plasma proteomic analysis identified biomarkers of COVID-19 disease progression [1]. The top pathways identified by Shu et al. were those of platelet degranulation and the complement and coagulation cascades [1]. These identified pathways were complementary to another recent study comparing COVID-19 disease and control subjects, where proteomic panels also identified biological pathways involved in platelet degranulation and the coagulation cascade [2]. Whilst the comparison with absolute disease-free normality is relevant, an increasing proportion of the population have insulin resistant states with associated metabolic conditions; an example of such a metabolic condition is polycystic ovary syndrome (PCOS) where it has been shown that protein expression patterns may differ compared to those without PCOS [3]. Notably, in PCOS, platelet aggregation enhancement together with aberrant diminished plasma fibrinolytic activity potentially giving rise to enhanced thrombosis

has been described [4, 5], with markers of coagulation being enhanced [6].

For a protein biomarker to be of value, there needs to be a clear discrimination between normal and disease condition levels. Therefore, platelet degranulation and the complement and coagulation cascade proteomic analysis was performed in women with and without PCOS to compare with these pathways described in COVID-19 disease [1].

243 subjects (146 PCOS and 97 control women) were recruited to the local PCOS biobank (ISRCTN70196169) [3] in the Department of Endocrinology, Hull and East Yorkshire Hospitals NHS Trust. The Rotterdam consensus diagnostic criteria were used to diagnose PCOS. Proteins that were described for platelet degranulation (18 of 27 proteins) and the complement and coagulation cascades (16 of 19 proteins) [1] were measured using the Slow Off-rate Modified Aptamer (SOMA)-scan plasma protein measurement [7], shown in Table 1. Statistics were performed using Graphpad Prism 8.0.

As reported previously [3], whilst cohorts were age-matched, the PCOS women differed in having increased systolic and diastolic blood pressure and waist circumference ($p < 0.05$), together with increased insulin resistance, increased androgens and C-reactive protein (CRP) ($p < 0.001$), indicative of metabolic dysfunction.

For the 46 protein biomarkers described by Shu et al. [1], 34 were available for measurement in the Somalogic platform: 4 of 18 were found to differ in PCOS for platelet

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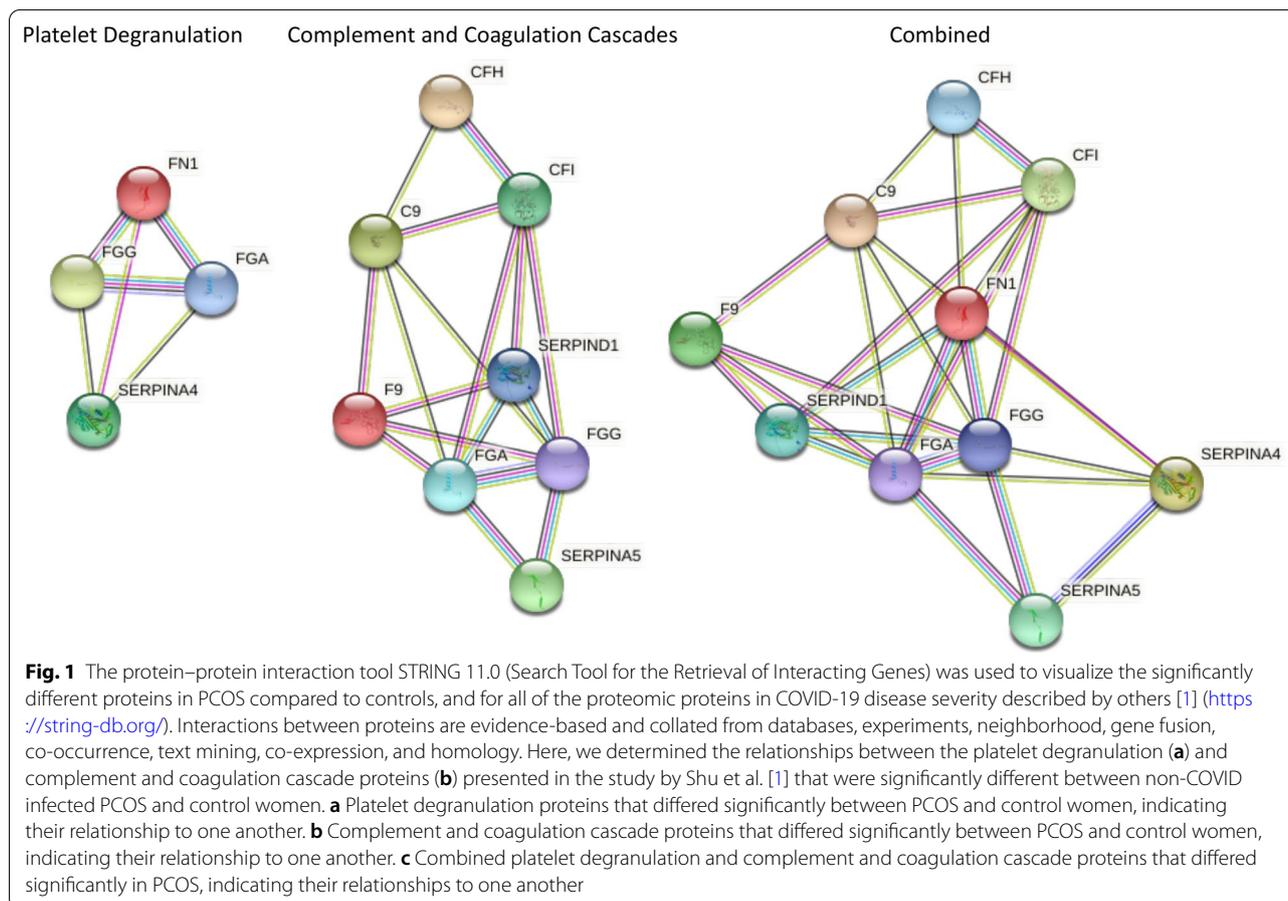


Table 1 Proteins identified as being altered in COVID-19 disease categorized according to **A**, platelet degranulation; **B**, complement and coagulation cascades in non-COVID infected polycystic ovary syndrome (PCOS) and control women

a PLATELET DEGRANULATION					b COMPLEMENT AND COAGULATION CASCADES				
Target Full Name	Target	UniProt	Entrez Gene Symbol	T-test Control vs PCOS	Target Full Name	Target	UniProt	Entrez Gene Symbol	T-test Control vs PCOS
Transgelin-2	Transgelin-2	P37802	TAGLN2	0.869	Fibrinogen gamma chain	Fibrinogen g-chain dimer	P02679	FGG	0.00002
Neutrophil-activating peptide 2	NAP-2	P02775	PPBP	0.274	Fibrinogen	Fibrinogen	P02671, P02675, P02679	FGA	0.013
Fibrinogen gamma chain	Fibrinogen g-chain dimer	P02679	FGG	0.00002	Complement factor I	Factor I	P05156	CFI	0.00000002
Fibrinogen	Fibrinogen	P02671, P02675, P02679	FGA	0.013	von Willebrand factor	vWF	P04275	VWF	0.077
Thrombospondin-1	Thrombospondin-1	P07996	THBS1	0.151	Coagulation factor IX	Coagulation Factor IX	P00740	F9	<0.00001
Platelet factor 4	PF-4	P02776	PF4	0.32	Complement factor H	Factor H	P08603	CFH	<0.000000001
von Willebrand factor	vWF	P04275	VWF	0.077	Complement component C9	C9	P02748	C9	0.00002
Serpin peptidase inhibitor, clade A (alpha-1 antitrypsin, antitrypsin), member 3	alpha-1-antichymotrypsin complex	P01011	SERPINA3	0.2672	Vitronectin	Vitronectin	P04004	VTN	0.393
Fructose-bisphosphate aldolase A	aldolase A	P04075	ALDOA	0.373	Plasminogen	Plasminogen	P00747	PLG	0.674
Metalloproteinase inhibitor 1	TIMP-1	P01033	TIMP1	0.165	Kininogen-1	"Kininogen, HMW"	P01042	KNG1	0.797
Plasminogen	Plasminogen	P00747	PLG	0.674	Alpha-2-antiplasmin	a2-Antiplasmin	P08697	SERPINF2	0.168
Kininogen-1	"Kininogen, HMW"	P01042	KNG1	0.797	Complement component C4B		P0C0L4, P0C0L5, P0C0L4, P0C0L5	C4B	0.687
Alpha-2-antiplasmin	a2-Antiplasmin	P08697	SERPINF2	0.168	Plasma serine protease inhibitor	PCI	P05154	SERPINA5	0.038
Fibronectin	Fibronectin	P02751	FN1	0.0098	Heparin cofactor 2	Heparin cofactor II	P05546	SERPIND1	0.00003
CD109 antigen	CD109	Q6YHK3	CD109	0.205	Vitamin K-dependent protein C	Protein C	P04070	PROC	0.511
Alpha-2-macroglobulin	a2-Macroglobulin	P01023	A2M	0.266	Alpha-2-macroglobulin	a2-Macroglobulin	P01023	A2M	0.266
Kallistatin	Kallistatin	P29622	SERPINA4	0.0006					
Apolipoprotein A-I	Apo A-I	P02647	APOA1	0.328					

Table 1 (continued)

Students' t-test was used to determine differences between protein levels. Proteins that differed significantly ($p < 0.05$) are shown in red font. Proteins that are common to both the platelet degranulation and the complement/coagulation cascades are shaded in orange



degranulation [fibrinogen-gamma chain ($p=0.00002$), fibrinogen ($p=0.013$), fibronectin ($p=0.0098$) and kallistatin ($p=0.0006$)], whilst 8 of 16 proteins for complement and coagulation cascade (fibrinogen-gamma chain ($p=0.00002$), fibrinogen ($p=0.013$), complement factor 1 ($p=0.00000002$), coagulation factor IX ($p < 0.00001$), complement factor H ($p < 0.000000001$), complement component C9 ($p=0.00002$), plasma serine protease inhibitor ($p=0.038$) and heparin cofactor 2 ($p=0.00003$) (Table 1). Moreover, those proteins that significantly differed between PCOS and controls share a close relationship to one another, as shown by the protein–protein interaction tool STRING (Search Tool for the Retrieval of Interacting Genes) pathways (Fig. 1).

The significant difference seen in PCOS compared to controls indicates the need for validation of such markers in the non-COVID-19 infected population before they can be considered as biomarkers for COVID-19 and its severity. Notably, based on the indication that COVID-19

severity can be related to these markers, their detection in a PCOS COVID-19 positive patient may give a false impression of severity, potentially leading to the introduction of inappropriate therapy; conversely, the detection of these specific markers in women with PCOS may actually indicate that more proactive intervention is required, as these women may have a propensity for increased COVID-19 disease severity [8].

Limitations of the study include that the Somalogic panel did not include all of the proteins that were previously reported, and the proteomic analysis differed so may not be directly comparable to the Shu et al. study [1] or others [2]. Nonetheless, the majority of proteins were common to both proteomic platforms.

In conclusion, 12 of 34 protein biomarkers contained within the platelet degranulation and complement and coagulation cascades and purported to indicate disease progression in patients infected with COVID-19, differed between non-COVID-19 infected PCOS and control

women. This indicates that validation of such proposed COVID-19 specific biomarkers is a necessity, although it is unclear if this places PCOS women at increased risk of more severe COVID-19 disease.

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Authors' contributions

ASMM, MN and AEB analyzed the data and wrote the manuscript. TS supervised clinical studies and edited the manuscript. SLA contributed to study design, data interpretation and the writing of the manuscript. All authors reviewed the final version of the manuscript. AEB is the guarantor of this work. All authors read and approved the final manuscript.

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Availability of data and materials

All the data for this study will be made available upon reasonable request to the corresponding author.

Ethics approval and consent to participate

The Newcastle & North Tyneside Ethics committee approved this study that was conducted according to the Declaration of Helsinki. All study participants signed an informed consent form prior to participation.

Consent for publication

All authors gave their consent for publication.

Competing interests

No authors have any conflict of interest or competing interests to declare.

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