

REVIEW

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Allo-priming as a universal anti-viral vaccine: protecting elderly from current COVID-19 and any future unknown viral outbreak

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Abstract

Background: We present the rationale for a novel allo-priming approach to serve the elderly as a universal anti-virus vaccine, as well serving to remodel the aging immune system in order to reverse immunosenescence and inflammaging. This approach has the potential to protect the most vulnerable from disease and provide society an incalculable economic benefit. Allo-priming healthy elderly adults is proposed to provide universal protection from progression of any type of viral infection, including protection against progression of the current outbreak of COVID-19 infection, and any future variants of the causative SARS-CoV-2 virus or the next 'Disease X'. Allo-priming is an alternative approach for the COVID-19 pandemic that provides a back-up in case vaccination strategies to elicit neutralizing antibody protection fails or fails to protect the vulnerable elderly population. The allo-priming is performed using activated, intentionally mismatched, ex vivo differentiated and expanded living Th1-like cells (AlloStim[®]) derived from healthy donors currently in clinical use as an experimental cancer vaccine. Multiple intradermal injections of AlloStim[®] creates a dominate titer of allo-specific Th1/CTL memory cells in circulation, replacing the dominance of exhausted memory cells of the aged immune system. Upon viral encounter, by-stander activation of the allo-specific memory cells causes an immediate release of IFN- γ , leading to development of an "anti-viral state", by-stander activation of innate cellular effector cells and activation of cross-reactive allo-specific CTL. In this manner, the non-specific activation of allo-specific Th1/CTL initiates a cascade of spatial and temporal immune events which act to limit the early viral titer. The release of endogenous heat shock proteins (HSP) and DAMP from lysed viral-infected cells, in the context of IFN- γ , creates of conditions for in situ vaccination leading to viral-specific Th1/CTL immunity. These viral-specific Th1/CTL provide sterilizing immunity and memory for protection from disease recurrence, while increasing the pool of Th1/CTL in circulation capable of responding to the next viral encounter.

Conclusion: Allo-priming has potential to provide universal protection from viral disease and is a strategy to reverse immunosenescence and counter-regulate chronic inflammation (inflammaging). Allo-priming can be used as an adjuvant for anti-viral vaccines and as a counter-measure for unknown biological threats and bio-economic terrorism.

Keywords: COVID-19, Immunosenescence, Inflammaging, Cell therapy, Immunotherapy, Vaccine

Background

Herein we propose to use a novel allo-priming strategy using patented, allogeneic Th1-like immune cells conjugated to CD3/CD28 microbeads (AlloStim[®]) to serve as a "Universal Anti-Viral Vaccine" to protect the health of elderly adults. Vaccination has been a strategy to protect against viral diseases in adults, such as influenza,

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pneumococcal pneumonia, shingles and hepatitis A/B. Successful prophylactic vaccination mechanisms provide protection through eliciting neutralizing antibodies to prevent viral entry into cells. However, this strategy does not provide protection against antigenic shift or drift variants of the original virus [1, 2]. Currently, there are at least three known variants of the SARS-CoV-2 virus [3].

In addition, pathological viruses are intracellular and not always accessible to antibodies. For this reason, neutralizing antibody vaccines have not been effective against a number of complex viruses, including HIV, HCV, CMV, Zika, RSV, Dengue and SARS/MERS. For the same reason, convalescent serum prophylaxis and treatment may not be able to confer sterilizing immunity or memory. These sophisticated viruses may require an effective cellular immune response for sterilizing immunity [4–8]. Without sterilizing cellular immunity, there can be viral recurrence as has been reported with COVID-19 [9].

Many efforts are underway to develop anti-viral vaccines which elicit protective cellular immunity [10], but these have not yet been successfully translated to demonstrate clinical benefit [11]. The age-related functional decline in cellular immunity (immunosenescence) makes the elderly less able to mount a cellular immune response to vaccination, making this population more vulnerable to morbidity and mortality associated with viral diseases and less likely to respond to an anti-viral vaccine.

In addition, elderly also suffer detrimental effects on their immune function due to chronic inflammation, known as “inflammaging” [12]. Inflammaging is correlated with comorbidities such as cancer, atherosclerosis, neurodegenerative diseases (e.g., Alzheimer’s and Parkinson’s disease) all which increase the likelihood of serious progression of viral infection. In addition, the aging of the structure and function of the lungs contributes to increased incidence of pneumonia, acute respiratory distress syndrome (ARDS) and sepsis in the elderly after respiratory viral infection.

The remodeling of the senescent immune systems of the elderly through allo-priming is proposed as a method to restore cellular immune function in this population. The ability to restore functional cellular immunity to the elderly can increase responsiveness to viral infections, including COVID-19 and any future emergent novel virus. In essence, an elderly immune system modulated by allo-priming would potentially respond to viral infection in a similar manner to the immune response of younger individuals, resulting in less serious disease. The immunomodulation of the elderly immune system to function more like a youthful immune system should also restore responsiveness to any current or future viral-specific vaccines. A more balanced immune system in the

elderly can also counter-regulate inflammaging, providing broad ranging health benefits to the elderly and to society [13].

Main text

Allopriming

The allo-priming concept is designed to prime the immune systems of healthy elderly adults to create high titers of allo-specific Th1/CTL memory cells which can become activated upon encounter with any virus (bystander activation) [14]. The by-stander activation of allo-specific Th1/CTL memory cells upon viral encounter causes the release of interferon-gamma (IFN- γ), which creates an “anti-viral state” [15]. IFNs create an antiviral state in both virus-infected cells and uninfected, bystander cells, by inducing a program of gene transcription that interferes with multiple stages of viral replication cycles through various mechanisms [16].

The IFN- γ release from by-stander activated allo-specific Th1/CTL memory cells activates innate effector cells (e.g., NK, NKT and macrophages) which in turn release additional IFN- γ , sustaining the anti-viral state. These activated innate effector cells lyse viral infected cells, controlling acute viral burden. By-stander activation of resident allo-specific CTL can also cross-react to lyse viral infected cells [17]. The lysis of viral infected cells by activated innate effector cells and cross-reactive allo-specific memory CTL releases “danger signals” [18] and heat shock proteins (HSP) [19] which chaperone viral antigens (e.g., GRP78, HSP70) [20, 21] into the microenvironment, creating the conditions for “in situ vaccination”, which leads to development of viral-specific cellular immunity.

Antigen presenting cells (APC), such as dendritic cells (DC), engulf and process released HSP-chaperoned viral antigens. Processing in the context of danger signals and IFN- γ , causes the maturation of immature DC to IL-12+ DC [22, 23]. IL-12 production acts to further increase IFN- γ , sustaining the anti-viral state and upregulating MHCI, MHCII and co-stimulatory molecules on APC [24, 25]. The mature DC migrate to regional lymph nodes and display viral antigens on MHC I and MHC II with co-stimulator CD80/86 expression, leading to viral-specific Th1/CTL expansion. The IFN- γ upregulates MHCI on viral infected cells so these cells can be recognized and killed by viral-specific CTL [26]. These viral-specific CTL can then orchestrate a sterilizing immune response and later differentiate into memory cells that provides protection from re-infection.

Thus, allo-antigenic priming can modulate the systemic immune balance and provide a pool of non-exhausted allo-specific Th1/CTL memory cells capable of rapidly responding to viral infection. The non-specific activation of these allo-specific memory cells upon viral encounter

causes release of IFN- γ . The release of IFN- γ orchestrates the sequential activation of immune cells resulting in formation of an antiviral state, innate elimination of invading viruses and development of a viral-specific effector response and memory.

Each time this cascade is initiated by viral encounter, the virus elimination serves as a booster to the original allo-antigenic priming. The new viral-specific Th1/CTL memory cells resulting from viral elimination join the resident memory allo-specific Th1/CTL resulting from the allo-antigenic priming, increasing the titer of Th1/CTL memory cells in circulation. Encounter with each new virus will non-specifically activate all previous non-exhausted memory cells and support the immune cascade that eliminates the new virus. Memory Th1/CTL to each new virus, in turn, increases the titer of Th1/CTL memory cells primed to respond to a subsequent viral encounter.

Creating a memory Th1/CTL immune response to alloantigen thus provides protection against an unrelated viral infection through the immune mechanism known as “heterologous immunity”. Heterologous immunity occurs when by-stander activation of immune memory cells to one virus alters the immune response to, and the course of infection of, an unrelated virus encountered later [27, 28].

In this manner, healthy adults can be provided pan-viral protection against viral infections, including SARS-CoV-2, influenza A/B and any future variants and unknown novel viruses that may emerge. This self-amplifying process can provide long-term disease mitigation and universal protection against known and unknown viral infections for the elderly.

Immunity to alloantigens

In order to effectuate the anti-viral and anti-aging immune mechanisms, the allo-priming must elicit allo-specific Th1/CTL immunity. Injection of alloantigen alone can elicit humoral and Th2 responses, and multiple injections can result in tolerance to the allo-antigens. Many factors, both intrinsic and extrinsic to the allograft, can influence the nature and magnitude of the allo-rejection immune response, including the nature of the allograft, the site of the body where it is placed, and the immunological status of the recipient [29].

To assure consistent differentiation of allo-specific Th1/CTL memory after allo-antigen priming, regardless of host immune status and age, a bioengineered, patented, immune cell called “AlloStim[®]” is used. AlloStim[®] is living, activated, intentionally mismatched, ex vivo expanded and differentiated allogeneic Th1-like cells derived from healthy donors. Injection of undifferentiated

allogeneic immune cells or non-activated allogeneic immune cells were not able to elicit allo-specific Th1 memory, while priming with activated allogeneic Th1-like AlloStim[®] cells elicited dominant Th1 immunity and had both a protective effect and a therapeutic effect in a cancer model [30, 31].

AlloStim[®] cells are administered intradermally (ID) in an activated state. To assure the injected cells are activated when administered, they are prepared with anti-CD3/anti-CD28-coated microbeads attached. The activated AlloStim[®] cells are formulated as a frozen dosage form at 1×10^7 cells/ml in PlasmaLyte A containing 1% human serum albumin and 2% DMSO. Individual doses of 0.5 ml are thawed and administered intradermally every 3–4 days for up to five total doses. This priming schedule is sufficient to provide high titers of circulating allo-specific Th1/CTL. AlloStim[®] has been evaluated in Phase I/II, Phase II and Phase IIA clinical trials in USA and Thailand in chemotherapy-refractory metastatic solid tumor patients. Phase IIB pre-registration and Phase IIB/III randomized, controlled registration clinical trials are currently being prepared for launch in third line metastatic colorectal cancer and advanced/metastatic hepatocellular carcinoma in USA and Asia, respectively. In these human clinical trials, AlloStim[®] has demonstrated ability to down-regulate checkpoint molecules in the tumor microenvironment, cause system-wide infiltration of effector T-cells and NK cells into metastatic lesions (conversion from “cold” to “hot”) and has consistently demonstrated a tail of between 20 and 35% of long-term survivors in chemotherapy-refractory metastatic disease with a good safety profile [32, 33].

Activated AlloStim[®] express high density CD40L and type 1 cytokines, including IFN- γ , TNF- α and GM-CSF. AlloStim[®] has been shown to modulate the immune systems of heavily pre-treated metastatic cancer patients (which resemble the senescent immune systems of the elderly) and has been used in protocols which were designed to elicit the same anti-tumor mechanism of allogeneic stem cell transplant procedures (graft vs tumor or “GVT”) without the need for a matched donor, chemotherapy conditioning or risk of GVHD [30, 31, 34–38].

Allogenic cells are highly immunogenic and are rejected even by immunocompromised hosts [39]. The rejection of the mis-matched cells eliminates risk of GVHD side-effects of allogeneic cell administration. Multiple ID injections of AlloStim[®] amplify the titer of allo-specific Th1/CTL cells in circulation and a portion of these cells differentiate into long lasting memory cells. These allo-specific Th1/CTL memory cells provide a functional pool of immune cells that are capable of non-specific (by-stander) activation [40] upon encounter with virus [14, 41] resulting in immediate release of IFN- γ .

The production of proinflammatory cytokines by AlloStim[®], including IFN- γ and TNF- α , initially activates host NK cells to reject the allogeneic cells, due to missing self-MHC I on the CD4+ AlloStim[®] cells. The NK cell compartment is highly stable in terms of function and phenotype in the elderly [42] and can therefore readily reject mismatched allogeneic cells. In subsequent ID AlloStim[®] injections, the production of IFN- γ and expression of CD40L by AlloStim[®] activates macrophages that reject the AlloStim[®] [43]. The rejection of the allogeneic cells by NK cells or macrophages in the skin results in release of endogenous HSP and DAMP danger signals [22], which in the context of CD40L and IFN- γ expressed by AlloStim[®] result in DC maturation to IL-12+ DC1 phenotype and their migration to draining lymph nodes to interact with cognate T-cells to elicit allo-specific Th1/CTL immunity [44].

Elderly adults

The current COVID-19 viral pandemic has led to high number of deaths worldwide. This is the third serious coronavirus (CoV) outbreak in less than 20 years, following SARS in 2002–2003 and MERS in 2012. Elderly people account disproportionately to the morbidity and mortality associated with CoV infection. Vaccines have been the main global strategy to minimize the impact of viral infections, but no successful vaccines have yet been developed for these previous highly virulent CoVs. In any event, elderly adults generally respond poorly to antiviral vaccines [45–47]. This creates a high unmet medical need for a vaccine to provide protection to the elderly from the current COVID-19 pandemic and protect from any future pandemics.

While healthy younger adults generally present with more mild symptoms in response to viral infection, the elderly are slow to respond and are susceptible to higher viral titers and chronic viral infection which leads to progression to severe symptoms, especially in the setting of respiratory viral infections [48–54]. The delayed and poorly effective responsiveness of elderly to viral infection is due, in large part, to compromised cellular immunity [55–57] related to “immunosenescence” [58–60].

Elderly individuals have a significantly delayed innate immune response to viral infection and the adaptive immune response that follows is often sub-optimal [61]. The delayed innate immune response results in accumulation of high viral titers which, in turn, serves to elicit a late over-active immune response which is correlated with a more severe and often lethal clinical course [62]. The acute high viral levels that are achieved in elderly as a result of late innate immune response also increases the virulence of the virus, by increasing chances of human-to-human spread of infection.

There is a dysregulation in the Th1/Th2-system in the elderly, which is dominated by Th2-functions [63, 64]. Memory Th1/CTL cells in elderly adults are diminished and functionally exhausted [65–68]. Memory cells of the elderly become exhausted due to lifelong antigenic challenge which can be as a result of chronic viral infection (e.g., CMV, EBV, HBV, HCV) [69, 70]. Exhausted memory cells are unable to be non-specifically activated upon viral encounter. This defect in cellular immunity in the elderly limits the availability of heterologous immune mechanisms to react to acute viral infection, which contributes to a slower and often inadequate innate response to viral infection.

Elderly also have limited T-cell repertoires [71, 72] which inhibits their ability to develop viral-specific adaptive T-cell immune responses, both prophylactically (by vaccination) [73], and in situ (as a result of adaptive immune clearance of infection). This suppressed cellular immunity together with the aging of the respiratory organs makes elderly more susceptible to pathological effects of viral infection, especially effects of respiratory viral infection [74]. The immune defects of the elderly make this population less likely to benefit from vaccinations as preventative measures against infectious diseases [75].

The current inability to provide elderly with protection from viral infection has an enormous economic impact on society [76, 77]. The medical and economic impact of pandemics which disproportionately affect the elderly will be exacerbated as the proportion of the adult population increases globally. The current projections indicate that by 2050, the group of 60 years and older will represent 21.1% of the population [78]. Therefore, there is an urgent unmet need to develop strategies to reverse immunosenescence and to develop methods which provide protection of the elderly population from viral epidemics.

Immunomodulation of the elderly immune system

The progressive age-related dysregulation and decline of the immune system is known as “immunosenescence”. In the aging immune system, there is an accumulation of memory cells as a result of chronic stimulation with repeated clinical and subclinical infections [13]. This chronic stimulation causes these memory cells to become exhausted and no longer able to function, contributing to an increased susceptibility of elderly to infectious pathogens [79, 80]. The aging immune system is also characterized by a decline in the numbers of naive T cells, an imbalance in Th1/Th2 cell subsets, and a decrease in T cell receptor (TCR) repertoire [75, 81, 82].

A cellular immune response with release of Th1 (type 1) cytokine profile, including IFN- γ , TNF- α and IL-2, is

known to be protective against most viral infections [83]. IFN- γ in particular has been shown to play an essential role in clearance of viral infection [84]. Modulating the immune system of elderly individuals through alloantigen priming to provide high titers of non-exhausted Th1/CTL memory cells that can be non-specifically activated upon encounter with any virus to cause release type 1 cytokines may provide an immediate anti-viral immune response upon viral exposure and could also reinstate responsiveness to viral vaccines [85]. The delay in IFN responses to virus infection in the elderly is due to active suppression by viral proteins and immunosenescence [86], which allows virus to reach high titers early in infection, leading to immunopathology, T-cell apoptosis and progression to lethal pneumonia [87, 88].

The immediate release of IFN- γ upon acute viral infection corrects the problem of delayed immune responsiveness and creates an early “anti-viral state” [89, 90]. Establishment of the anti-viral state with early release of IFN- γ provides a crucial initial line of defense against viral infection [91]. The anti-viral state is normally induced through early release of type I/III IFN from viral infected cells [92]. However, many viruses, including coronavirus, can suppress production of type I/III IFN as an immune avoidance mechanism [93–96]. The strategy of providing a pool of non-exhausted memory cells which produce Type II IFN (IFN- γ) upon viral encounter, overcomes the viral escape. IFN- γ creates an anti-viral state even in type I/III IFN resistant cells [97]. Further, IFN- γ mobilizes innate effector cells that can respond rapidly to eliminate viral infection [98] and also produce IFN- γ . Thus, early IFN- γ response to viral infection through allo-priming provides a rapid means of control over viral infection in the elderly.

Chronic inflammation in the elderly is known to be associated with increased susceptibility to cancer [99, 100], atherosclerosis [101] and neurodegenerative disorders [102, 103]. The modulation of the elderly immune system that occurs by allo-priming can also have the effect of counter-regulating resident chronic inflammation [104, 105] which causes “inflammaging” resulting in reversing of immunosenescence [106]. Thus, allo-priming protocols may not only provide universal anti-viral protection for the elderly, but may also serve as an “anti-aging” mechanism to protect from co-morbid diseases of aging.

Clinical features of viral infection in elderly

Allo-priming is predicted to reduce the severity of clinical symptoms upon encounter with virus. Clinical features of respiratory viral infections in humans vary from a first subset that experience mild flu-like symptoms which subside over a few days; to a second subset that

experience moderate to severe flu-like symptoms associated with high fever, hypoxemia and progression to pneumonia-like symptoms. These symptomatic patients are at high risk to progress to acute respiratory distress syndrome (ARDS), sepsis, multiple organ failure and death. The subset with mild flu-like symptoms is associated with younger healthy individuals, while the subset associated with more severe symptoms is mostly associated with elderly adults that are frail and/or present with co-morbidities.

The progression to more serious symptoms can occur even when there is a decline in virus titers [107] connected to a late over-exuberant immune response. The initial delayed immune response leads to high viral titers which is followed by an exaggerated late immune response and a “cytokine storm”. This sequence of events is believed to be responsible for the severe symptoms related to respiratory virus infections in the elderly.

During pathogenic virus infection, a cytokine storm leads to excessive inflammatory infiltrates and virus-induced tissue destruction which contributes to morbidity and mortality. The timing and source of the IFN in the cytokine storm can affect the clinical course of viral disease. High serum levels of IFN- γ late in the course of viral infection is correlated with more severe disease [108] and immunopathology [109], whereas early IFN release results in lower viral titers and less severe disease. These observations support the prediction that early activation and release of IFN- γ from non-exhausted allo-specific memory Th1/CTL will correct this problem and avoid progression to severe disease.

The type of cytokine storm associated with severe disease is caused by cytokines released by activated monocytes, producing a storm of IFN- α , CCL-2, IL-6, TNF- α , CCL-3, CCL-5, CxCL-2, IL-1 α , and IFN- γ . This monokine cytokine storm is directly correlated with morbidity and mortality [110]. The majority of fatalities associated with cytokine storm also developed bacterial pneumonia and sepsis. It is recognized that a major cause of respiratory failure is coexistent bacterial pneumonia leading to ARDS. ARDS is characterized by damage to the endothelial–epithelial barrier of the alveoli, resulting in fluid leakage and accumulation in the alveolar lumen inhibiting gas exchange.

The elderly have a pre-disposition to develop progressive respiratory compromise and sepsis. In fact, sepsis is considered a disease of aging [111] and is among the top causes of ICU admissions of the elderly. The features of sepsis-induced immunosuppression, independent of age, share many of the same characteristics of immunosenescence. During the last decades, there has been a significant increase in incidence of sepsis in patients over 65 years of age [112]. Many respiratory virus patients that

progress to develop pneumonia and ARDS [113, 114] later die due to sepsis.

The overactive immune response to sepsis, including monokine “cytokine storm” results from over activated M1 macrophages in response to tissue damage [115]. The response to sepsis is similar between old and younger patients. However, the mortality is much higher in older patients [116]. The difference is related to the dysregulated immune systems of the elderly. The elderly are unable to turn down the over-activated monocyte response, leading to disease progression, while regulatory mechanisms in the young counter-act and prevent the consequences of monocyte over-activation.

The dysregulated immune system of the elderly is related to the increased activity of myeloid-derived suppressor cells (MDSC) [117, 118]. MDSCs are potent inducers of immunosenescence [119], sepsis-acquired immunodeficiency [120] as well as cancer metastasis and progression [121]. MDSC are positively correlated with IL-6 and negatively correlated with IFN- γ [122, 123].

Immunosenescence causes an imbalanced homeostatic regulation of innate and adaptive immune responses, diminishing the host capacity to rapidly restore balanced immune functions. Thus, combined effects of age-induced immunosuppression, delayed innate immune responses [86], exaggerated late immune responses due to altered homeostasis [88], all combine to make the post-viral infection period in the elderly have a longer duration. As a result, there are increased levels of viral propagation, higher incidences of tissue injury, and increased progression to ARDS and septic shock in the elderly.

Accordingly, allo-priming enabling the early release of IFN- γ in response to viral infection can prevent the differentiation of MDSC and counter-regulate resident MDSC immunosuppression in the elderly [124]. This will have the effect of remodeling the elderly immune system in a manner which will prevent the severe symptoms of viral infection and progression to ARDS and sepsis.

Conclusion

Highly pathogenic human CoVs pose a substantial threat to public health. There are basically two groups of patients upon exposure to CoV, with the majority developing a short duration of clinical symptoms and the minority experiencing severe disease characterized by pneumonia and ARDS. The elderly are disproportionately vulnerable to severe disease due to immunosenescence and comorbidities. It is likely that CoVs will continue to cross species and cause additional outbreaks in the future. Therefore, even if a vaccine is developed for prevention of COVID-19, it would not protect against the next outbreak of a novel virus. Therefore, there is a need

to develop novel strategies, not only to control the current pandemic, but also to be prepared to prevent the next pandemic.

Strategies to develop a vaccine to elicit neutralizing antibodies to SARS-CoV-2 are technically challenging due to the conformational hiding of the receptor binding domain (RBD) and the ability of this virus to transfer cell-to-cell in syncytia and infect target cells in a ACE2- and protease-independent manner by pinocytosis, limiting environmental exposure needed for antibody neutralization activity. Development of an anti-viral vaccine against complex viruses, such a SARS-Cov-2, is likely to require a potent and broad T-cell response to overcome viral escape mechanisms related to humoral immunity.

A vaccine that elicits a robust cellular immune response requires identification of conserved viral epitopes and effective processing and presentation of viral antigens by APC on MHC I and MHC II in conjunction with co-stimulatory signals, as well as a diversity in the naive T cell repertoire. In addition, CTL memory cells resulting from vaccination will require infected cells to present the selected viral antigens in the vaccine on MHC I. Selection of common viral epitopes which present on MHC I (and MHC II) for vaccine development is technically difficult and CoV infection causes the down-regulation of MHC I on infected cells, making infected cells invisible to CTL. Since the elderly lack naive T-cells that are necessary for development of CTL with broad specificity to viral antigens, elderly would be less likely to respond efficiently to any future vaccine which targets cellular immunity.

The rejection of allografts is one of the most conserved and powerful immune mechanisms, making priming with alloantigens an ideal candidate for use in vaccination of the elderly. AlloStim[®] is in use under a US FDA cleared IND and has been shown to readily be rejected by heavily pre-treated, immunosuppressed metastatic cancer patients. A phase I/II clinical trial protocol for use of AlloStim[®] in healthy elderly adults is currently under review by US FDA. Upon clearance, clinical trials could be initiated to evaluate this allo-priming concept in short order. For proof-of-concept, longitudinal PBMC samples are proposed to be collected pre- and post-allo-priming. The PBMC are to be pulsed with a panel of inactivated viruses or recombinant viral proteins to determine if they will non-specifically activate the allo-specific Th1/CTL memory cells. The supernatants from the viral-pulsed PBMC are proposed to be evaluated in live virus cytolytic plaque assays to determine if viral propagation is suppressed.

Elderly persons are particularly susceptible to progressive viral disease and have delayed immune responses to viral infection due, at least in part, to immunosenescence,

Th2 immune bias, decreased diversity of naïve T-cells and high frequencies of exhausted memory cells from chronic inflammation (e.g., CMV). This results in a natural loss of the ability to mount effective innate and adaptive cellular immune responses to invading pathogens. The delayed and suppressed cellular immune response in the elderly enables viruses to become widely established and the resulting increased viral titers causes dys-regulated immune responses that can lead to a late, over-active immune response which can progress to pneumonitis, multiple organ failure and death.

The key to an effective natural innate immune response that can limit the course of virus infection is the early production of type I/III interferons (IFN) and subsequent switch to type II IFN (IFN- γ) production as the immune response matures. Successful viral infections become established due, in large part, to delayed innate immune responses and viral-mediated suppression of type I/III IFN production, allowing for rapid viral propagation which eventually results in dys-regulated immune responses which are correlated with tissue destruction, co-infection, multiple organ failure and death.

To prevent accumulation of high viral burden in the elderly upon viral infection, allo-priming provides a mechanism whereby a ready pool of de-novo primed T-cells are in circulation that can respond rapidly to viral infection by producing IFN- γ . The presence of non-exhausted Th1/CTL memory immune cells will modify the elderly immune character by providing a Th1 re-balancing mechanism in the memory cell compartment. This is accomplished through the creation of a high titer of polyclonal, allo-specific, non-exhausted, memory Th1/CTL T-cells through intradermal injections of activated allogeneic Th1-like cells (AlloStim[®]). The allo-specific memory cells resulting from the priming are programmed to produce IFN- γ upon activation. IFN- γ has a direct anti-viral effect on cells infected with virus and can also protect uninfected cells from infection. IFN- γ creates the same anti-viral environment as innate release of Type I/III IFN does in an effective natural innate immune response.

These allo-specific memory cells are capable of cross-reacting with foreign viral antigens and can be readily activated non-specifically by environmental stimuli such as cytokines and foreign pathogens. The memory pool of allo-specific immune cells in primed individuals can also be re-activated by additional intradermal or intravenous infusion of AlloStim[®] allogeneic cells upon first symptoms of viral disease. The by-stander effect of allo-specific Th1/CTL memory T-cell activation and IFN- γ production is predicted to elicit protective effects on cells in the respiratory tract and generate rapid immune-mediated

viral clearance as well as condition the microenvironment in infected tissues (e.g., lung epithelial cells) for an in situ vaccination leading to viral-specific immunity and memory that is specific for the invading virus. The heterologous immune effect can amplify the pan-viral protection upon each viral encounter, providing the possibility of long-term protection.

The proposed alloantigen priming strategy can also be used in conjunction with any viral-specific vaccines. Co-injection of allo-specific and viral specific antigens could accelerate the adaptive cellular immune response to a known virus and serve as a means to enhance response to vaccines in the elderly. Subsequent injection of allogeneic cells will stimulate an allo-rejection memory recall response which can non-specifically activate resident viral-specific cells elicited by vaccination. This dual activation provides a mechanism to upregulate MHC I on infected targets, making the memory cells elicited by viral-specific vaccination able to identify and kill viral infected targets. The combination approach also has the advantage of overcoming the narrow immunity conferred by a single peptide vaccine by incorporating the in situ vaccination mechanism to elicit broad viral-specific cellular immune responses.

This allo-priming mechanism can also be used as a counter-measure to bio-terrorism and bio-economic terrorism. Individuals that have been primed with alloantigen could be treated with an emergency injection of alloantigen to activate innate immunity and initiate the cascade of immune events leading to clearance of an unknown pathogen. Alloantigen could be provided in syringes for emergency ID injection upon concern for bioweapon exposure or first occurrence of symptoms, much in the same manner as epinephrine is provided to prevent anaphylactic shock.

While finding methods to treat and prevent COVID19 infection is of urgent priority, it is also important to consider that the current pandemic is the third major outbreak of a novel coronaviral infection in humans within the past 20 years. Currently there are no registered vaccines or means of therapeutic protection against the prior SARS or MERS outbreaks available anywhere in the world. This is despite considerable efforts from experts worldwide to develop vaccines. The same technical obstacles preventing development of specific vaccines and treatments for these past CoV outbreaks likely also exist for development of a vaccine for the current COVID-19 viral pandemic. However, even if an effective vaccine is developed, it is estimated that it will take at least a year to complete clinical testing sufficient to obtain regulatory approval. If COVID-19 fades as did SARS and MERS, by the time a vaccine is available it will no longer be as urgent.

However, protection of the vulnerable elderly population from the next pandemic and protecting the world economy from natural and bioweapon threats will remain an urgent need. Therefore, novel approaches to addressing the problem of new emerging viral epidemics, especially in the vulnerable elderly population, are of a high priority. Allo-priming holds promise as such an approach.

Abbreviations

ARDS: Acute respiratory distress syndrome; CMV: Cytomegalovirus; CoV: Coronavirus; CTL: Cytotoxic T-lymphocyte; CCL: C–C motif ligand; CxCL: CxCL motif ligand; DAMP: Danger associated molecular pattern; DC: Dendritic cell; FDA: Food and Drug Administration; GM-CSF: Granulocyte-macrophage colony stimulating factor; GRP: Glucose-regulated protein; GVHD: Graft vs. host disease; GVT: Graft vs. tumor; HIV: Human immunodeficiency virus; HCV: Hepatitis C virus; HSP: Heat shock protein; ID: Intradermal; IFN: Interferon; IL: Interleukin; IND: Investigational New Drug; MDSC: Myeloid-derived suppressor cell; MERS: Middle East Respiratory Syndrome; MHC: Major histocompatibility complex; NK: Natural killer cell; NKT: Natural killer T cell; US: United States; SARS: Severe acute respiratory syndrome; Th1: T-helper 1; TNF: Tumor necrosis factor.

Acknowledgements

Not applicable.

Authors' contributions

MHN conceived of the allopriming concept and wrote the manuscript. RO reviewed and edited the manuscript. Both authors read and approved the final manuscript.

Funding

Not applicable.

Availability of data and materials

Not applicable.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

MHN is the Founder of Immunovative Therapies, Ltd. and Mirror Biologics, Inc which own patent rights to AlloStim®.

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Received: 21 April 2020 Accepted: 4 May 2020

Published online: 12 May 2020

References

- Operschall E, Pavlovic J, Nawrath M, Molling K. Mechanism of protection against influenza A virus by DNA vaccine encoding the hemagglutinin gene. *Intervirology*. 2000;43:322–30.
- Carrat F, Flahault A. Influenza vaccine: the challenge of antigenic drift. *Vaccine*. 2007;25:6852–62.
- Forster P, Forster L, Renfrew C, Forster M. Phylogenetic network analysis of SARS-CoV-2 genomes. *Proc Natl Acad Sci USA*. 2020;117:9241–3.
- Oxenius A, Price DA, Hersberger M, Schlaepfer E, Weber R, Weber M, Kundig TM, Boni J, Joller H, Phillips RE, et al. HIV-specific cellular immune response is inversely correlated with disease progression as defined by decline of CD4+ T cells in relation to HIV RNA load. *J Infect Dis*. 2004;189:1199–208.
- Nakamura I, Imawari M. Cellular immune response in HCV infection. *J Gastroenterol*. 2000;35:881–9.
- Kash JC, Muhlberger E, Carter V, Grosch M, Perwitasari O, Proll SC, Thomas MJ, Weber F, Klenk HD, Katze MG. Global suppression of the host antiviral response by Ebola- and Marburgviruses: increased antagonism of the type I interferon response is associated with enhanced virulence. *J Virol*. 2006;80:3009–20.
- Cassaniti I, Percivalle E, Adzasehoun KMG, Comolli G, Baldanti F. Dengue virus-specific humoral and T cellular immune response in Italian residents and travelers returning from endemic areas. *Vector Borne Zoonotic Dis*. 2020;20:295–302.
- Chen J, Lau YF, Lamirande EW, Paddock CD, Bartlett JH, Zaki SR, Subbarao K. Cellular immune responses to severe acute respiratory syndrome coronavirus (SARS-CoV) infection in senescent BALB/c mice: CD4+ T cells are important in control of SARS-CoV infection. *J Virol*. 2010;84:1289–301.
- Chen D, Xu W, Lei Z, Huang Z, Liu J, Gao Z, Peng L. Recurrence of positive SARS-CoV-2 RNA in COVID-19: a case report. *Int J Infect Dis*. 2020;93:297–9.
- Panagiotti E, Klenerman P, Lee LN, van der Burg SH, Arens R. Features of effective T cell-inducing vaccines against chronic viral infections. *Front Immunol*. 2018;9:276.
- Clemens EB, van de Sandt C, Wong SS, Wakim LM, Valkenburg SA. Harnessing the power of T cells: the promising hope for a universal influenza vaccine. *Vaccines*. 2018;6(2):18. <https://doi.org/10.3390/vaccines6020018>.
- Kirkwood KL. Inflammaging. *Immunol Invest*. 2018;47:770–3.
- Ventura MT, Casciaro M, Gangemi S, Buquicchio R. Immunosenescence in aging: between immune cells depletion and cytokines up-regulation. *Clin Mol Allergy*. 2017;15:21.
- Dosch SF, Mahajan SD, Collins AR. SARS coronavirus spike protein-induced innate immune response occurs via activation of the NF-kappaB pathway in human monocyte macrophages in vitro. *Virus Res*. 2009;142:19–27.
- Horvath CM, Darnell JE Jr. The antiviral state induced by alpha interferon and gamma interferon requires transcriptionally active Stat1 protein. *J Virol*. 1996;70:647–50.
- Yan N, Chen ZJ. Intrinsic antiviral immunity. *Nat Immunol*. 2012;13:214–22.
- Cornberg M, Clute SC, Watkin LB, Saccoccio FM, Kim SK, Naumov YN, Brehm MA, Aslan N, Welsh RM, Selin LK. CD8 T cell cross-reactivity networks mediate heterologous immunity in human EBV and murine vaccinia virus infections. *J Immunol*. 2010;184:2825–38.
- Gallucci S, Matzinger P. Danger signals: SOS to the immune system. *Curr Opin Immunol*. 2001;13:114–9.
- Breloer M, Dorner B, More SH, Roderian T, Fleischer B, von Bonin A. Heat shock proteins as "danger signals": eukaryotic Hsp60 enhances and accelerates antigen-specific IFN-gamma production in T cells. *Eur J Immunol*. 2001;31:2051–9.
- Wati S, Soo ML, Zilm P, Li P, Paton AW, Burrell CJ, Beard M, Carr JM. Dengue virus infection induces upregulation of GRP78, which acts to chaperone viral antigen production. *J Virol*. 2009;83:12871–80.
- Menoret A. Purification of recombinant and endogenous HSP70s. *Methods*. 2004;32:7–12.
- Zhang X, Mosser DM. Macrophage activation by endogenous danger signals. *J Pathol*. 2008;214:161–78.
- Fournier P, Arnold A, Schirrmacher V. Polarization of human monocyte-derived dendritic cells to DC1 by in vitro stimulation with Newcastle disease virus. *J BUON*. 2009;14(Suppl 1):S111–22.
- Berberich C, Ramirez-Pineda JR, Hambrecht C, Alber G, Skeiky YA, Moll H. Dendritic cell (DC)-based protection against an intracellular pathogen is dependent upon DC-derived IL-12 and can be induced by molecularly defined antigens. *J Immunol*. 2003;170:3171–9.
- Ahuja SS, Mummidi S, Malech HL, Ahuja SK. Human dendritic cell (DC)-based anti-infective therapy: engineering DCs to secrete functional IFN-gamma and IL-12. *J Immunol*. 1998;161:868–76.
- Yang Y, Xiang Z, Ertl HC, Wilson JM. Upregulation of class I major histocompatibility complex antigens by interferon gamma is necessary

- for T-cell-mediated elimination of recombinant adenovirus-infected hepatocytes in vivo. *Proc Natl Acad Sci USA*. 1995;92:7257–61.
27. Welsh RM, Che JW, Brehm MA, Selin LK. Heterologous immunity between viruses. *Immunol Rev*. 2010;235:244–66.
 28. Messina NL, Zimmermann P, Curtis N. The impact of vaccines on heterologous adaptive immunity. *Clin Microbiol Infect*. 2019;25:1484–93.
 29. Marino J, Paster J, Benichou G. Allorecognition by T lymphocytes and allograft rejection. *Front Immunol*. 2016;7:582.
 30. Har-Noy M, Zeira M, Weiss L, Fingerut E, Or R, Slaviv S. Allogeneic CD3/CD28 cross-linked Th1 memory cells provide potent adjuvant effects for active immunotherapy of leukemia/lymphoma. *Leuk Res*. 2009;33:525–38.
 31. Har-Noy M, Zeira M, Weiss L, Slaviv S. Completely mismatched allogeneic CD3/CD28 cross-linked Th1 memory cells elicit anti-leukemia effects in unconditioned hosts without GVHD toxicity. *Leuk Res*. 2008;32:1903–13.
 32. Har-Noy M, Sukepainsarnjaroen W, Lausoontornsiri W, Wittenberg A, Bloch M, Sonnenfeld T, Fingerut E, Kaminer Y, Ahojja A, Wongwiwatchai J, et al. Individualized cancer vaccine to elicit tumor debulking immunity in advanced HCC. *J Clin Oncol*. 2018;36:e15054.
 33. Har-Noy M, Lausoontornsiri W, Or R, Katsanis E. Response of HER2+ breast cancer patients to allogeneic cell immunotherapy. *J Clin Oncol*. 2012;30:e13013.
 34. Epple LM, Bemis LT, Cavanaugh RP, Skope A, Mayer-Sonnenfeld T, Frank C, Olver CS, Lencioni AM, Dusto NL, Tal A, et al. Prolonged remission of advanced bronchoalveolar adenocarcinoma in a dog treated with autologous, tumour-derived chaperone-rich cell lysate (CRCL) vaccine. *Int J Hyperth*. 2013;29:390–8.
 35. Mayer-Sonnenfeld T, Har-Noy M, Lillehei KO, Graner MW. Proteomic analyses of different human tumour-derived chaperone-rich cell lysate (CRCL) anti-cancer vaccines reveal antigen content and strong similarities amongst the vaccines along with a basis for CRCLs unique structure: CRCL vaccine proteome leads to unique structure. *Int J Hyperth*. 2013;29:520–7.
 36. LaCasse CJ, Janikashvili N, Larmonier CB, Alizadeh D, Hanke N, Kartchner J, Situ E, Centuori S, Har-Noy M, Bonnotte B, et al. Th-1 lymphocytes induce dendritic cell tumor killing activity by an IFN-gamma-dependent mechanism. *J Immunol*. 2011;187:6310–7.
 37. Janikashvili N, LaCasse CJ, Larmonier C, Trad M, Herrell A, Bustamante S, Bonnotte B, Har-Noy M, Larmonier N, Katsanis E. Allogeneic effector/memory Th-1 cells impair FoxP3+ regulatory T lymphocytes and synergize with chaperone-rich cell lysate vaccine to treat leukemia. *Blood*. 2011;117:1555–64.
 38. Har-Noy M, Slaviv S. The anti-tumor effect of allogeneic bone marrow/stem cell transplant without graft vs. host disease toxicity and without a matched donor requirement? *Med Hypotheses*. 2008;70:1186–92.
 39. Richters CD, Hoekstra MJ, du Pont JS, Kreis RW, Kamperdijk EW. Immunology of skin transplantation. *Clin Dermatol*. 2005;23:338–42.
 40. Bangs SC, Baban D, Cattani HJ, Li CK, McMichael AJ, Xu XN. Human CD4+ memory T cells are preferential targets for bystander activation and apoptosis. *J Immunol*. 2009;182:1962–71.
 41. Eberl G, Brawand P, MacDonald HR. Selective bystander proliferation of memory CD4+ and CD8+ T cells upon NK T or T cell activation. *J Immunol*. 2000;165:4305–11.
 42. Manser AR, Uhrberg M. Age-related changes in natural killer cell repertoires: impact on NK cell function and immune surveillance. *Cancer Immunol Immunother*. 2016;65:417–26.
 43. Liu W, Xiao X, Demirci G, Madsen J, Li XC. Innate NK cells and macrophages recognize and reject allogeneic nonself in vivo via different mechanisms. *J Immunol*. 2012;188:2703–11.
 44. Benichou G, Yamada Y, Yun SH, Lin C, Fray M, Tocco G. Immune recognition and rejection of allogeneic skin grafts. *Immunotherapy*. 2011;3:757–70.
 45. Ongradi J, Kovesi V. Factors that may impact on immunosenescence: an appraisal. *Immun Ageing*. 2010;7:7.
 46. Targonski PV, Jacobson RM, Poland GA. Immunosenescence: role and measurement in influenza vaccine response among the elderly. *Vaccine*. 2007;25:3066–9.
 47. Ongradi J, Stercz B, Kovesi V, Vertes L. Immunosenescence and vaccination of the elderly. I. Age-related immune impairment. *Acta Microbiol Immunol Hung*. 2009;56:199–210.
 48. Falsey AR, Walsh EE. Respiratory syncytial virus infection in elderly adults. *Drugs Aging*. 2005;22:577–87.
 49. Falsey AR, Hennessey PA, Formica MA, Cox C, Walsh EE. Respiratory syncytial virus infection in elderly and high-risk adults. *N Engl J Med*. 2005;352:1749–59.
 50. Saxena AK, Panhotra BR. The vulnerability of middle-aged and elderly patients to hepatitis C virus infection in a high-prevalence hospital-based hemodialysis setting. *J Am Geriatr Soc*. 2004;52:242–6.
 51. Butt AA, Dascomb KK, DeSalvo KB, Bazzano L, Kissinger PJ, Szerlip HM. Human immunodeficiency virus infection in elderly patients. *South Med J*. 2001;94:397–400.
 52. Marcus EL, Tur-Kaspa R. Epidemiology of hepatitis C virus infection in the elderly. *Gerontology*. 2001;47:117–8.
 53. Brunner S, Herndler-Brandstetter D, Weinberger B, Grubeck-Loebenstein B. Persistent viral infections and immune aging. *Ageing Res Rev*. 2011;10:362–9.
 54. Haq K, McElhaney JE. Ageing and respiratory infections: the airway of ageing. *Immunol Lett*. 2014;162:323–8.
 55. Pawelec G, Adibzadeh M, Solana R, Beckman I. The T cell in the ageing individual. *Mech Ageing Dev*. 1997;93:35–45.
 56. Katz AE. Immunity and aging. *Otolaryngol Clin N Am*. 1982;15:287–92.
 57. Gonzalez PA, Bueno SM, Riedel CA, Kalergis AM. Impairment of T cell immunity by the respiratory syncytial virus: targeting virulence mechanisms for therapy and prophylaxis. *Curr Med Chem*. 2009;16:4609–25.
 58. Ma Y, Fang M. Immunosenescence and age-related viral diseases. *Sci China Life Sci*. 2013;56:399–405.
 59. Bektas A, Schurman SH, Sen R, Ferrucci L. Human T cell immunosenescence and inflammation in aging. *J Leukoc Biol*. 2017;102:977–88.
 60. Pera A, Campos C, Lopez N, Hassouneh F, Alonso C, Tarazona R, Solana R. Immunosenescence: implications for response to infection and vaccination in older people. *Maturitas*. 2015;82:50–5.
 61. Sun Y, Lopez CB. The innate immune response to RSV: advances in our understanding of critical viral and host factors. *Vaccine*. 2017;35:481–8.
 62. Li G, Fan Y, Lai Y, Han T, Li Z, Zhou P, Pan P, Wang W, Hu D, Liu X, et al. Coronavirus infections and immune responses. *J Med Virol*. 2020;92:424–32.
 63. Rink L, Cakman I, Kirchner H. Altered cytokine production in the elderly. *Mech Ageing Dev*. 1998;102:199–209.
 64. Shearer GM. Th1/Th2 changes in aging. *Mech Ageing Dev*. 1997;94:1–5.
 65. Deng Y, Jing Y, Campbell AE, Gravenstein S. Age-related impaired type 1 T cell responses to influenza: reduced activation ex vivo, decreased expansion in CTL culture in vitro, and blunted response to influenza vaccination in vivo in the elderly. *J Immunol*. 2004;172:3437–46.
 66. Karanflov CI, Liu B, Fox CC, Lakshmanan RR, Whisler RL. Age-related defects in Th1 and Th2 cytokine production by human T cells can be dissociated from altered frequencies of CD45RA+ and CD45RO+ T cell subsets. *Mech Ageing Dev*. 1999;109:97–112.
 67. Akbar AN, Fletcher JM. Memory T cell homeostasis and senescence during aging. *Curr Opin Immunol*. 2005;17:480–5.
 68. Sansoni P, Vescovini R, Fagnoni F, Biasini C, Zanni F, Zanlari L, Telera A, Lucchini G, Passeri G, Monti D, et al. The immune system in extreme longevity. *Exp Gerontol*. 2008;43:61–5.
 69. Saeidi A, Zandi K, Cheok YY, Saeidi H, Wong WF, Lee CYQ, Cheong HC, Yong YK, Larsson M, Shankar EM. T-cell exhaustion in chronic infections: reversing the state of exhaustion and reinvigorating optimal protective immune responses. *Front Immunol*. 2018;9:2569.
 70. Henson SM, Akbar AN. Memory T-cell homeostasis and senescence during aging. *Adv Exp Med Biol*. 2010;684:189–97.
 71. Minato N, Hattori M, Hamazaki Y. Physiology and pathology of T-cell ageing. *Int Immunol*. 2020;32:223–31.
 72. Lepletier A, Alsharif A, Chidgey AP. Inflammation and thymus ageing. *Front Horm Res*. 2017;48:19–36.
 73. Dugan HL, Henry C, Wilson PC. Aging and influenza vaccine-induced immunity. *Cell Immunol*. 2020;348:103998.
 74. Sharma G, Goodwin J. Effect of aging on respiratory system physiology and immunology. *Clin Interv Aging*. 2006;1:253–60.
 75. Oh SJ, Lee JK, Shin OS. Aging and the immune system: the impact of immunosenescence on viral infection, immunity and vaccine immunogenicity. *Immune Netw*. 2019;19:e37.
 76. Szucs T. The socio-economic burden of influenza. *J Antimicrob Chemother*. 1999;44(Suppl B):11–5.

77. Maynard A, Bloor K. The economic impact of pandemic influenza. *BMJ*. 2009;339:b4888.
78. Roses M, Bonvehi PE. Vaccines in adults. *Medicina (B Aires)*. 2019;79:552–8.
79. Crooke SN, Ovsyannikova IG, Poland GA, Kennedy RB. Immunosenescence and human vaccine immune responses. *Immun Ageing*. 2019;16:25.
80. Hakim FT, Flomerfelt FA, Boyiadzis M, Gress RE. Aging, immunity and cancer. *Curr Opin Immunol*. 2004;16:151–6.
81. Muller L, Di Benedetto S, Pawelec G. The immune system and its dysregulation with aging. *Subcell Biochem*. 2019;91:21–43.
82. Sadighi Akha AA. Aging and the immune system: an overview. *J Immunol Methods*. 2018;463:21–6.
83. Zhu J, Yamane H, Paul WE. Differentiation of effector CD4 T cell populations (*). *Annu Rev Immunol*. 2010;28:445–89.
84. Stark GR, Kerr IM, Williams BR, Silverman RH, Schreiber RD. How cells respond to interferons. *Annu Rev Biochem*. 1998;67:227–64.
85. Wagner A, Garner-Spitzer E, Jasinska J, Kollaritsch H, Stiasny K, Kundi M, Wiedermann U. Age-related differences in humoral and cellular immune responses after primary immunisation: indications for stratified vaccination schedules. *Sci Rep*. 2018;8:9825.
86. Lau SKP, Lau CCY, Chan KH, Li CPY, Chen H, Jin DY, Chan JFW, Woo PCY, Yuen KY. Delayed induction of proinflammatory cytokines and suppression of innate antiviral response by the novel Middle East respiratory syndrome coronavirus: implications for pathogenesis and treatment. *J Gen Virol*. 2013;94:2679–90.
87. Channappanavar R, Fehr AR, Vijay R, Mack M, Zhao J, Meyerholz DK, Perlman S. Dysregulated type I interferon and inflammatory monocyte-macrophage responses cause lethal pneumonia in SARS-CoV-infected mice. *Cell Host Microbe*. 2016;19:181–93.
88. Channappanavar R, Perlman S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. *Semin Immunopathol*. 2017;39:529–39.
89. Kang S, Brown HM, Hwang S. Direct antiviral mechanisms of interferon-gamma. *Immune Netw*. 2018;18:e33.
90. Shrestha B, Wang T, Samuel MA, Whitby K, Craft J, Fikrig E, Diamond MS. Gamma interferon plays a crucial early antiviral role in protection against West Nile virus infection. *J Virol*. 2006;80:5338–48.
91. Levy DE, Garcia-Sastre A. The virus battles: IFN induction of the antiviral state and mechanisms of viral evasion. *Cytokine Growth Factor Rev*. 2001;12:143–56.
92. Gaajetaan GR, Geelen TH, Vernooy JH, Dentener MA, Reynaert NL, Rohde GG, Beuken EV, Grauls GE, Bruggeman CA, Stassen FR. Interferon-beta induces a long-lasting antiviral state in human respiratory epithelial cells. *J Infect*. 2013;66:163–9.
93. Aizaki H, Saito S, Ogino T, Miyajima N, Harada T, Matsuura Y, Miyamura T, Kohase M. Suppression of interferon-induced antiviral activity in cells expressing hepatitis C virus proteins. *J Interferon Cytokine Res*. 2000;20:1111–20.
94. Guo P. Suppression of interferon-mediated antiviral immunity by hepatitis B virus: an overview of research progress. *Scand J Immunol*. 2013;78:230–7.
95. Comar CE, Goldstein SA, Li Y, Yount B, Baric RS, Weiss SR. Antagonism of dsRNA-induced innate immune pathways by NS4a and NS4b accessory proteins during MERS coronavirus infection. *mBio*. 2019;10:e00319-19. <https://doi.org/10.1128/mBio.00319-19>.
96. Siu KL, Chan CP, Kok KH, Chiu-Yat Woo P, Jin DY. Suppression of innate antiviral response by severe acute respiratory syndrome coronavirus M protein is mediated through the first transmembrane domain. *Cell Mol Immunol*. 2014;11:141–9.
97. Romeo G, Affabris E, Federico M, Mechti N, Coccia EM, Jemma C, Rossi GB. Establishment of the antiviral state in alpha, beta-interferon-resistant Friend cells treated with gamma-interferon. Induction of 67-kilodalton protein kinase activity in absence of detectable 2-5A synthetase. *J Biol Chem*. 1985;260:3833–8.
98. Anderson MJ. The role of interferon in the NK cell killing of virus-infected target cells. *J Hyg*. 1982;89:347–51.
99. Pawelec G. Immunosenescence and cancer. *Biogerontology*. 2017;18:717–21.
100. Zanussi S, Serraino D, Dolcetti R, Berretta M, De Paoli P. Cancer, aging and immune reconstitution. *Anticancer Agents Med Chem*. 2013;13:1310–24.
101. Yu HT, Park S, Shin EC, Lee WW. T cell senescence and cardiovascular diseases. *Clin Exp Med*. 2016;16:257–63.
102. Costantini E, D'Angelo C, Reale M. The role of immunosenescence in neurodegenerative diseases. *Mediat Inflamm*. 2018;2018:6039171.
103. Solana C, Tarazona R, Solana R. Immunosenescence of natural killer cells, inflammation, and alzheimer's disease. *Int J Alzheimers Dis*. 2018;2018:3128758.
104. Whary MT, Fox JG. Th1-mediated pathology in mouse models of human disease is ameliorated by concurrent Th2 responses to parasite antigens. *Curr Top Med Chem*. 2004;4:531–8.
105. Strober W, Kelsall B, Fuss I, Marth T, Ludviksson B, Ehrhardt R, Neurath M. Reciprocal IFN-gamma and TGF-beta responses regulate the occurrence of mucosal inflammation. *Immunol Today*. 1997;18:61–4.
106. Witkowski JM, Bryl E, Fulop T. Should we try to alleviate immunosenescence and inflammaging—why, how and to what extent? *Curr Pharm Des*. 2019;25:4154–62.
107. Peiris JS, Chu CM, Cheng VC, Chan KS, Hung IF, Poon LL, Law KI, Tang BS, Hon TY, Chan CS, et al. Clinical progression and viral load in a community outbreak of coronavirus-associated SARS pneumonia: a prospective study. *Lancet*. 2003;361:1767–72.
108. Chien JY, Hsueh PR, Cheng WC, Yu CJ, Yang PC. Temporal changes in cytokine/chemokine profiles and pulmonary involvement in severe acute respiratory syndrome. *Respirology*. 2006;11:715–22.
109. Cameron MJ, Ran L, Xu L, Danesh A, Bermejo-Martin JF, Cameron CM, Muller MP, Gold WL, Richardson SE, Poutanen SM, et al. Interferon-mediated immunopathological events are associated with atypical innate and adaptive immune responses in patients with severe acute respiratory syndrome. *J Virol*. 2007;81:8692–706.
110. Oldstone MB, Rosen H. Cytokine storm plays a direct role in the morbidity and mortality from influenza virus infection and is chemically treatable with a single sphingosine-1-phosphate agonist molecule. *Curr Top Microbiol Immunol*. 2014;378:129–47.
111. Monneret G, Gossez M, Venet F. Sepsis and immunosenescence: closely associated in a vicious circle. *Aging Clin Exp Res*. 2019. <https://doi.org/10.1007/s40520-019-01350-z>.
112. Reinhart K, Daniels R, Kissonoo N, Machado FR, Schachter RD, Finfer S. Recognizing sepsis as a global health priority—a WHO resolution. *N Engl J Med*. 2017;377:414–7.
113. Bauer TT, Ewig S, Rodloff AC, Muller EE. Acute respiratory distress syndrome and pneumonia: a comprehensive review of clinical data. *Clin Infect Dis*. 2006;43:748–56.
114. Joynt GM, Yap HY. SARS in the intensive care unit. *Curr Infect Dis Rep*. 2004;6:228–33.
115. Isobe KI, Nishio N, Hasegawa T. Immunological aspects of age-related diseases. *World J Biol Chem*. 2017;8:129–37.
116. Kale SS, Yende S. Effects of aging on inflammation and hemostasis through the continuum of critical illness. *Aging Dis*. 2011;2:501–11.
117. Alves AU, Bueno V. Immunosenescence: participation of T lymphocytes and myeloid-derived suppressor cells in aging-related immune response changes. *Einstein*. 2019;17:eRB4733.
118. Hurez V, Daniel BJ, Sun L, Liu AJ, Ludwig SM, Kioussis MJ, Thibodeaux SR, Pandeswara S, Murthy K, Livi CB, et al. Mitigating age-related immune dysfunction heightens the efficacy of tumor immunotherapy in aged mice. *Cancer Res*. 2012;72:2089–99.
119. Salminen A, Kaamiranta K, Kauppinen A. Immunosenescence: the potential role of myeloid-derived suppressor cells (MDSC) in age-related immune deficiency. *Cell Mol Life Sci*. 2019;76:1901–18.
120. Venet F, Demaret J, Gossez M, Monneret G. Myeloid cells in sepsis-acquired immunodeficiency. *Ann N Y Acad Sci*. 2020. <https://doi.org/10.1111/nyas.14333>.
121. Netherby CS, Abrams SI. Mechanisms overseeing myeloid-derived suppressor cell production in neoplastic disease. *Cancer Immunol Immunother*. 2017;66:989–96.
122. Gonda K, Shibata M, Ohtake T, Matsumoto Y, Tachibana K, Abe N, Ohto H, Sakurai K, Takenoshita S. Myeloid-derived suppressor cells are increased and correlated with type 2 immune responses, malnutrition, inflammation, and poor prognosis in patients with breast cancer. *Oncol Lett*. 2017;14:1766–74.
123. Nam S, Kang K, Cha JS, Kim JW, Lee HG, Kim Y, Yang Y, Lee MS, Lim JS. Interferon regulatory factor 4 (IRF4) controls myeloid-derived suppressor cell (MDSC) differentiation and function. *J Leukoc Biol*. 2016;100:1273–84.

124. Medina-Echeverez J, Haile LA, Zhao F, Gamrekelashvili J, Ma C, Metais JY, Dunbar CE, Kapoor V, Manns MP, Korangy F, Greten TF. IFN-gamma regulates survival and function of tumor-induced CD11b⁺ Gr-1^{high} myeloid derived suppressor cells by modulating the anti-apoptotic molecule Bcl2a1. *Eur J Immunol*. 2014;44:2457–67.

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