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Association of adiponectin gene variants with idiopathic recurrent miscarriage according to obesity status: a case–control study

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Abstract

Background: This study addresses whether the association of adiponectin gene (*ADIPOQ*) variants with idiopathic recurrent pregnancy loss (RPL) is influenced by obesity.

Methods: Retrospective case–control study performed in outpatient obstetrics/gynecology clinics. Study subjects comprised 308 women with RPL, defined as ≥ 3 consecutive miscarriages of unknown etiology, and 310 control women. *ADIPOQ* genotyping was done by allele exclusion method on real-time PCR.

Results: Of the 14 *ADIPOQ* variants tested, the minor allele frequency (MAF) of rs4632532, rs17300539, rs266729, rs182052, rs16861209, and rs7649121 were significantly higher, while rs2241767, and rs1063539 MAF were lower in RPL cases, hence assigning RPL-susceptibility and protection to these variants, respectively. Higher frequencies of heterozygous rs17300539 and rs16861209, and homozygous rs4632532, rs266729, and rs182052 genotypes, and reduced frequencies of heterozygous rs1063539 and rs2241767, homozygous rs2241766 genotypes were seen in RPL cases. *ADIPOQ* rs4632532, and rs2241766 were associated with RPL in obese, while rs1063539 and rs16861209 were associated with RPL in non-obese women; rs182052 and rs7649121 associated with RPL independently of BMI changes. Based on LD pattern, two haplotype blocks were identified. Within Block 1 containing rs4632532, rs16861194, rs17300539, rs266729, rs182052, rs16861209, rs822396, and rs7649121, increased frequency of CAGGACAT and TAACGAAA, and reduced frequency of TAGCGCAA haplotypes were seen in RPL cases when compared to controls, thereby assigning RPL susceptibility and protection, respectively.

Conclusion: This is the first study to document contribution of *ADIPOQ* variants and haplotypes with RPL, and also to underscore the contribution of obesity to genetic association studies.

Keywords: Adiponectin, Haplotypes, Idiopathic recurrent miscarriage, Real-time PCR

Background

Recurrent pregnancy loss (RPL), defined as ≥ 2 idiopathic miscarriages with the same partner prior the 20th week of gestation [1], is a common pregnancy complication which affects ~1–2% of women worldwide [2]. Modifiable and non-modifiable factors that contribute to RPL pathogenesis were reported. The former includes environmental and lifestyle factors (smoking, obesity),

poorly-controlled diabetes, hypothyroidism, and infections [3, 4], while the latter includes chromosomal anomalies, age at first pregnancy, and presence of specific genetic polymorphisms [2, 5]. Despite the identification of several risk factors of RPL, the etiology of idiopathic RPL remains not completely understood, with more than 50% of RPL cases still remain unexplained [1, 2].

Normal human pregnancy is linked with altered metabolism stemming from the accumulation of white adipose tissues [6], leading to progressive decrease in insulin sensitivity [6, 7]. White adipose tissues synthesize an array of hormones within the intra-uterine compartment, which serve both metabolic, anti-atherosclerotic, and

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anti-inflammatory roles. These include leptin, resistin, apelin, and adiponectin [8]. Adiponectin is a 30 kDa, 244 amino acid adipokine, which regulates insulin sensitivity, blood glucose levels, and lipid metabolism, and possesses anti-atherosclerosis and anti-inflammatory activities [9].

Adiponectin is produced mostly from adipocyte, but is also secreted by other tissues, including reproductive organs [9]. Adiponectin is detected in amniotic fluid after 15 weeks of pregnancy, and its maternal plasma concentrations remain virtually unchanged throughout pregnancy [10, 11]. Reduction in adiponectin levels results in reproductive disorders [12, 13], including failed embryo implantation, polycystic ovarian syndrome (PCOS), endometriosis, gestational diabetes mellitus (GDM) [14, 15], and RPL [16]. Adiponectin is encoded by *ADIPOQ* gene (adipocyte C1Q and collagen domain), located on chromosome 3q27 spanning 17 kB, and containing 3 exons and two introns [17], and is highly polymorphic. Previous studies demonstrated association of functional *ADIPOQ* gene variants with altered circulating adiponectin concentrations [18, 19], and with pregnancy complications, including GDM, metabolic syndrome, and preeclampsia (PE) [20–22]. Decreased adiponectin serum levels were also linked with severe obesity [23], endometriosis and PCOS [12].

Very few studies examined the association of *ADIPOQ* variants with idiopathic RPL. An earlier Indian study documented the association of *ADIPOQ* rs2241766 (exon 2) and rs15012299 (intron 2) with increased risk of RPL [16]. In addition, *ADIPOQ* rs2241766 and rs15012299 variants showed weak, but statistically significant, haplotype association with PE susceptibility in Finnish women [24]. Here we analyze the association of 14 common *ADIPOQ* variants in women with confirmed RPL diagnosis, and multiparous control women who were matched to RPL cases according to self-declared racial background. The contribution of these variants to RPL was examined according to obesity, and was examined at the allele, genotype, and haplotype levels.

Subjects and methods

Subjects

This retrospective case–control study was performed between January 2013 and June 2015. Study population included 308 women with confirmed RPL who were recruited consecutively from the outpatient OB/GYN clinics in Manama and Riffa (Bahrain). In addition, we recruited 310 multiparous women with two or more successful pregnancies as controls. The study protocol was approved by Arabian Gulf University Research and Ethics Committee (IRB approval: 35-PI-01/15), and was done according to Helsinki II Declaration. All patients provided informed written consent before blood sampling.

We defined RPL as ≥ 3 consecutive pregnancy losses of undetermined etiology, which occurred between the 7th–20th week of gestation, and with the same partner. RPL assessment was according to the guidelines of the Royal College of Obstetricians and Gynecologists (<https://www.rcog.org.uk/guidelines>). These required screening for LAC (lupus anticoagulant) and ACL (anticardiolipin) anti-phospholipid antibodies, and the inherited thrombophilias Factor V-Leiden (R506Q) and factor II/prothrombin G20210A. Karyotyping of both parents, and pelvic ultrasound scan by hysteroscopy or sonohysteroscopy for evaluation of uterine anatomy were also performed on all RPL cases. RPL cases were excluded if they were 40 years or older at first pregnancy, incompatibility in Rh blood groups, history of PE, which was defined as elevation in systolic/diastolic blood pressure (BP) exceeding 145/95 mmHg, or increase in systolic/diastolic BP exceeding 30/15 mmHg on at least two occasions, as well as biochemical pregnancy, and preclinical miscarriages. Cases were also excluded if they had systemic autoimmunity, thyroid dysfunction, diabetes, liver function abnormalities, anatomical disorders, and infections (toxoplasmosis, HIV, HCMV, Group B streptococci, *Chlamydia trachomatis*, hepatitis B and C viruses, rubella, and bacterial vaginosis).

Control women comprised hospital and university students and employees, and volunteers from the community, and were included if they had two or more successful pregnancies. Control women were included following check-up after uncomplicated pregnancy/delivery. Controls were excluded if they reported spontaneous and/or induced miscarriages, and family history of miscarriage, and were matched to RPL cases according to age ($P=0.94$), and self-reported ethnic origin (all Bahraini Arabs). Peripheral blood samples (2–5 ml) were collected from RPL cases and control women in EDTA-containing tubes for DNA extraction.

ADIPOQ genotyping

ADIPOQ single nucleotide polymorphisms (SNP) with minor allele frequencies (MAF) that exceeded 5% in Caucasians (HapMap CEU), were selected using SNPbrowser software 4.0 (ABI-ThermoFisher, Foster City, CA). Genotyping was done by the allelic discrimination method using VIC-/FAM-labelled primers (Table 2). Assay-on-demand TaqMan assays were obtained from ABI-ThermoFisher. PCR was performed in 6 μ l volume on StepOne Plus real-time PCR system, according to instructions of the manufacturer (ABI-ThermoFisher). A typical RT-PCR consisted of adding 2.2 μ l DNA template, and 4.0 μ l TaqMan genotype master mix (TaqMan 2X mix, 1.875 μ l nuclease free water, and 0.125 μ l 40X SNP primer mix) (Applied Biosystem). Pre-PCR (hold step) stage was

performed for 30 s at 60 °C, and for 10 min at 95 °C. The cycles conditions will be repeated 35 times, and consisted of denaturation (92 °C for 15 s), annealing and extension (60 °C for 1 min), followed by post-PCR stage at 60 °C for 30 s. Replicate quality control samples (\approx 10% of cases and controls) were included for assessment of genotyping reproducibility; concordance consistently exceeded 99%.

Statistical analysis

Statistical analysis was done on SPSS v. 24 (IBM, Armonk, NY). Categorical variables were expressed as percent of total, while continuous data were presented as mean \pm SD. Differences in means were analyzed using Student's t test, and inter-group significance was assessed by Pearson χ^2 or Fisher's exact test. Hardy-Weinberg equilibrium (HWE) calculation was done for control women using Haploview 4.2 (<https://www.broad.mit.edu/mpg/haploview>). SNPStats (<https://bioinformatics.iconologia.net/snpstats>) was used for genetic association analysis, under the assumption of additive genetic model. Power calculation for detection of association between *ADIPOQ* variants and RPL was done using CaTS Power Calculator (<https://www.sph.umich.edu/csg/abecasis/cats>). We used the following parameters: 308 RPL cases and 310 control women, relative risk for heterozygous and minor allele homozygous genotypes, and MAF for the 14 tested *ADIPOQ* SNPs RPL in cases and controls, and assuming a 2.5% population RPL prevalence in Bahrain (Bahrain Ministry of Health unpublished statistics). The overall power was calculated as 72.1%, which represented the average power of the included SNPs. Haploview 4.2 was used for determination of linkage disequilibrium (LD) and haplotype reconstruction, the latter done according to expectation maximization (EM) algorithm. Taking control women as the reference group, logistic regression analysis was used in determining odds ratios (OR) and 95% confidence intervals (CI) associated with the risk of RPL; $P < 0.05$ was taken as statistically significant.

Results

Demographic and clinical characteristics of RPL cases and control women

Table 1 summarizes the demographic and clinical profiles of RPL cases and control women. Age at entry into the study, fasting plasma glucose, gravida, and number of smokers were not significantly different between women with RPL and control subjects. Significant differences between the two study groups were seen in mean BMI ($P = 0.004$), menarche ($P < 0.001$), and in systolic/diastolic blood pressure readings ($P < 0.001$). While these did not constitute established RPL risk factors, we nevertheless

Table 1 Demographics and clinical characteristics of cases and controls

	Cases ^a	Controls ^a	P^b
Age at inclusion in study ^c	31.6 \pm 5.4	31.6 \pm 4.9	0.94
Body-mass index (kg/m ²) ^c	26.3 \pm 5.4	25.2 \pm 4.3	0.004
Obesity [n (%)] ^d	58 (19.6)	37 (12.1)	0.02
Smokers [n (%)] ^d	30 (10.1)	32 (10.8)	0.69
Systolic blood pressure (mmHg) ^c	114.2 \pm 11.9	120.2 \pm 17.0	<0.001
Diastolic blood pressure (mmHg) ^c	72.0 \pm 8.4	75.8 \pm 9.1	<0.001
Glucose (mmol/L) ^c	5.1 \pm 0.9	5.2 \pm 0.7	0.55
Menarche (years) ^c	12.2 \pm 1.1	12.8 \pm 1.0	<0.001
Number of pregnancies ^c	4.2 \pm 1.5	4.0 \pm 1.1	0.11
Number of children ^c	0.8 \pm 1.1	4.0 \pm 1.1	<0.001
Miscarriages ^c	3.6 \pm 1.0	0.0 \pm 0.1	<0.001

^a A total of 308 RPL cases and 310 control women were included

^b Student's t-test (continuous variables), Pearson's χ^2 test (categorical variables)

^c Mean \pm SD

^d Percent of total within each group/subgroup

included them as covariates which we controlled for in later analysis.

Association between *ADIPOQ* SNP and the risk of RPL

Table 2 presents the association between *ADIPOQ* SNP and RPL in women with RPL and control women. Genotype distributions of all 14 tested *ADIPOQ* variants conformed to HWE among study subjects. Among *ADIPOQ* SNP tested, higher MAF of rs4632532 ($P = 8.00 \times 10^{-3}$), rs17300539 ($P = 0.011$), rs266729 ($P = 4.00 \times 10^{-3}$), rs182052 ($P = 7.00 \times 10^{-4}$), rs16861209 ($P = 9.00 \times 10^{-3}$), rs7649121 ($P = 0.044$), thereby assigning RPL susceptibility to these variants. On the other hand, MAF of rs2241767 ($P = 0.041$) and rs1063539 ($P = 0.048$), was lower in women with RPL than in control women, suggesting RPL-protection associated with these *ADIPOQ* variants. No significant differences in MAF of the remaining SNPs were seen between women with RPL and controls.

Table 3 lists the distribution of *ADIPOQ* genotypes between women with RPL and control women. Taking major allele homozygous genotype (1/1) as the reference group (OR = 1.00), after controlling for BMI, systolic/diastolic blood pressure, and menarche, significantly higher frequencies of heterozygous rs17300539 (0.17 vs. 0.08) and rs16861209 (0.32 vs. 0.15), and homozygous rs4632532 (0.13 vs. 0.08), rs266729 (0.11 vs. 0.05), and rs182052 (0.16 vs. 0.05) genotypes were seen in RPL cases vs. control women. In addition, reduced genotype frequencies of heterozygous rs1063539 (0.21 vs. 0.31) and rs2241767 (0.22 vs. 0.31), and homozygous rs2241766 (0.02 vs. 0.07) were seen in women with RPL compared

Table 2 ADIPOQ SNPs analyzed in RPL cases and control women

SNP	Assay ID	Position	HWE P	Alleles	Cases ^a	Controls ^a	χ^2	P	aOR ^b (95% CI)	Power
<i>rs4632532</i>	C_27867233_10	186551682	0.209	T:C	0.351	0.271	7.035	0.008	1.46 (1.10–1.93)	90
<i>rs16861194</i>	C_33187775_10	186559425	0.325	A:G	0.098	0.079	0.979	0.322	1.25 (0.80–1.95)	63
<i>rs17300539</i>	C_33187774_10	186559460	1.000	G:A	0.087	0.044	6.534	0.011	2.06 (1.17–3.63)	100
<i>rs266729</i>	C_2412786_10	186559474	0.213	C:G	0.262	0.173	8.295	0.004	1.69 (1.18–2.42)	83
<i>rs182052</i>	C_2412785_10	186560782	0.348	G:A	0.360	0.249	11.381	7.00×10^{-4}	1.70 (1.25–2.31)	100
<i>rs16861209</i>	C_33187764_10	186563114	0.205	C:A	0.144	0.087	6.852	0.009	1.77 (1.15–2.72)	89
<i>rs822396</i>	C_2910316_10	186566877	0.337	A:G	0.139	0.157	0.582	0.445	0.92 (0.64–1.33)	42
<i>rs7649121</i>	C_42772949_10	186568785	0.303	A:T	0.182	0.134	4.063	0.044	1.44 (1.01–2.05)	
<i>rs2241766</i>	C_26426077_10	186570892	0.578	T:G	0.156	0.193	2.166	0.141	0.80 (0.57–1.12)	62
<i>rs1501299</i>	C_7497299_10	186571123	0.212	C:A	0.324	0.307	0.343	0.558	1.09 (0.83–1.43)	54
<i>rs2241767</i>	C_26426076_10	186571196	1.000	A:G	0.139	0.188	4.161	0.041	0.70 (0.49–0.98)	66
<i>rs3774261</i>	C_27479710_10	186571559	0.303	G:A	0.462	0.495	0.989	0.320	0.87 (0.67–1.14)	61
<i>rs6773957</i>	C_1486294_10	186573705	0.285	G:A	0.506	0.475	0.951	0.330	1.13 (0.88–1.46)	63
<i>rs1063539</i>	C_1486290_10	186575392	0.277	G:C	0.147	0.196	3.898	0.048	0.70 (0.50–0.99)	68

Italicface indicates statistical significance

^a MAF frequency

^b aOR = adjusted OR; variables that were controlled for were BMI, menarche, systolic and diastolic blood pressure

Table 3 ADIPOQ genotype frequencies

SNP	1/1 ^a			1/2 ^a			2/2 ^a		
	Cases	Controls	P ^b	Cases	Controls	OR (95% CI)	Cases	Controls	OR (95% CI)
<i>rs4632532</i>	141 (0.46) ^c	171 (0.55)	0.042	126 (0.41)	114 (0.37)	1.33 (0.92–1.93)	41 (0.13)	25 (0.08)	2.03 (1.11–3.70)
<i>rs1063539</i>	229 (0.74)	199 (0.64)	0.060	68 (0.21)	95 (0.31)	0.63 (0.41–0.96)	11 (0.04)	16 (0.05)	0.57 (0.23–1.44)
<i>rs17300539</i>	256 (0.83)	283 (0.91)	0.027	51 (0.17)	25 (0.08)	2.22 (1.22–4.04)	1 (0.003)	1 (0.003)	1.08 (0.07–17.41)
<i>rs16861209</i>	229 (0.74)	259 (0.84)	0.032	69 (0.32)	48 (0.15)	1.64 (1.01–2.68)	10 (0.03)	3 (0.01)	3.87 (0.79–18.88)
<i>rs266729</i>	187 (0.61)	208 (0.67)	0.079	88 (0.29)	86 (0.28)	1.14 (0.74–1.77)	33 (0.11)	16 (0.05)	2.37 (1.09–5.18)
<i>rs822396</i>	233 (0.76)	223 (0.72)	0.650	66 (0.21)	78 (0.25)	0.81 (0.53–1.25)	9 (0.03)	9 (0.03)	0.96 (0.33–2.80)
<i>rs1501299</i>	144 (0.47)	161 (0.52)	0.511	127 (0.41)	115 (0.37)	1.23 (0.85–1.79)	37 (0.12)	34 (0.11)	1.23 (0.69–2.17)
<i>rs182052</i>	132 (0.43)	174 (0.56)	0.001	128 (0.42)	119 (0.38)	1.43 (0.94–2.17)	48 (0.16)	17 (0.05)	3.67 (1.73–7.76)
<i>rs7649121</i>	222 (0.72)	244 (0.79)	0.190	57 (0.19)	49 (0.16)	1.30 (0.80–2.10)	29 (0.09)	17 (0.05)	1.76 (0.86–3.58)
<i>rs2241766</i>	219 (0.71)	195 (0.63)	0.043	82 (0.27)	96 (0.31)	0.77 (0.52–1.13)	7 (0.02)	22 (0.07)	0.34 (0.13–0.90)
<i>rs3774261</i>	92 (0.30)	93 (0.30)	0.212	148 (0.48)	127 (0.41)	1.19 (0.76–1.86)	68 (0.22)	90 (0.29)	0.78 (0.47–1.30)
<i>rs16861194</i>	257 (0.83)	264 (0.85)	0.687	46 (0.15)	44 (0.14)	1.09 (0.67–1.76)	5 (0.02)	2 (0.01)	2.01 (0.36–11.08)
<i>rs6773957</i>	80 (0.26)	88 (0.28)	0.790	144 (0.47)	144 (0.46)	1.11 (0.73–1.69)	84 (0.27)	78 (0.25)	1.18 (0.73–1.90)
<i>rs2241767</i>	223 (0.72)	200 (0.65)	0.054	67 (0.22)	96 (0.31)	0.62 (0.42–0.93)	18 (0.06)	14 (0.05)	1.14 (0.51–2.56)

Italicface indicates statistical significance

^a Genotypes were coded as per "1" = major allele, "2" = minor allele

^b 2-way ANOVA

^c Number of subjects (frequency)

to controls. The distribution of the remaining genotypes was not significantly different between women with RPL and control subjects.

Association of ADIPOQ polymorphisms with RPL in obese and non-obese RPL subjects

In view of the effect of ADIPOQ SNPs on adiponectin secretion, and the impact of adiposity of pregnancy outcome, we examined the association of the tested ADIPOQ polymorphisms with RPL in obese and non-obese RPL cases and controls. Women with RPL and control

Table 4 Association of ADIPOQ variants with RPL in obese vs. non-obese cases and control women

SNP	Non-obese				Obese			
	Control ^a	Case ^a	P	OR (95% CI)	Control ^a	Case ^a	P	OR (95% CI)
rs4632532	28.05	33.33	0.093	1.28 (0.96–1.71)	12.00	36.17	0.002	4.15 (1.60–10.76)
rs1063539	20.45	14.40	0.028	0.65 (0.44–0.95)	23.08	15.85	0.296	0.63 (0.26–1.51)
rs17300539	03.72	06.91	0.053	1.92 (0.98–3.75)	09.26	17.11	0.202	2.02 (0.68–6.06)
rs16861209	07.88	12.64	0.034	1.69 (1.03–2.76)	13.46	23.68	0.152	1.99 (0.77–5.19)
rs266729	19.17	24.00	0.117	1.33 (0.93–1.91)	16.67	30.26	0.125	2.17 (0.79–5.92)
rs822396	16.09	14.89	0.647	0.91 (0.62–1.35)	13.79	08.75	0.348	0.60 (0.20–1.76)
rs1501299	29.01	31.86	0.371	1.14 (0.85–1.54)	32.14	35.56	0.671	1.16 (0.57–2.36)
rs182052	26.90	37.50	0.004	1.63 (1.17–2.27)	10.71	31.43	0.005	3.82 (1.42–10.24)
rs7649121	13.78	19.19	0.040	1.49 (1.02–2.18)	10.71	26.09	0.046	2.60 (0.99–6.80)
rs2241766	20.62	16.09	0.093	0.74 (0.52–1.05)	30.00	14.44	0.021	0.39 (0.18–0.88)
rs3774261	32.08	45.45	0.386	0.88 (0.65–1.18)	46.00	48.68	0.764	1.11 (0.54–2.28)
rs16861194	08.63	09.05	0.823	1.05 (0.66–1.68)	00.00	07.45	–	N/A
rs6773957	49.53	51.50	0.571	1.08 (0.82–1.42)	41.38	46.94	0.498	1.25 (0.65–2.42)
rs2241767	19.82	15.79	0.125	0.76 (0.53–1.08)	24.14	19.39	0.484	0.76 (0.35–1.65)

Italicface indicates statistical significance

^a Percent minor allele carriers

women were sub grouped into non-obese and obese according to BMI cutoff of 30 kg/m². Results from Table 4 demonstrate differential association of ADIPOQ SNPs with RPL according to obesity was seen, with rs16861209 ($P=0.034$), rs182052 ($P=0.004$), and rs7649121 ($P=0.040$) being positively associated, while rs1063539 ($P=0.028$) was negatively associated with RPL in obese subjects (Table 3). In contrast, rs182052 ($P=0.005$), and rs7649121 ($P=0.046$) were positively, while rs4632532 ($P=0.002$), and rs2241766 ($P=0.021$) were negatively associated with RPL in obese subjects.

Identification of ADIPOQ haplotypes associated with RPL

The interaction between tested ADIPOQ variants, and their mode of inheritance in women with RPL and control women was next evaluated. The interaction between pair of SNP visualized by Haploview (Fig. 1). Haploview analysis showed marked LD among the tested ADIPOQ variants (Fig. 1), and defined two haploblocks. Haploblock 1 spanned 17 kb, and contained rs4632532, rs16861194, rs17300539, rs266729, rs182052, rs16861209, rs822396, and rs7649121, while Haploblock 2 spanned 4 kb, and contained rs2241766, rs1501299, rs2241767, rs3774261, rs6773957, and rs1063539.

“Common haplotype” was defined as the haplotype with frequencies >2% of the total haplotypes. Within Haploblock 1, 94.8% of 8-locus haplotype diversity was captured by 9 of the possible 256 haplotypes. Higher frequencies of CAGGACAT ($P=5.1 \times 10^{-3}$) and TAA-GAAA ($P=0.012$), and reduced frequency of TAGCGCAA ($P=7.6 \times 10^{-5}$) haplotypes were seen in RPL cases,

suggesting RPL-susceptible and RPL-protective aspect of these haplotypes, respectively. The frequencies of remaining haplotypes in Haploblock 1, and all haplotypes in Haploblock 2 were comparable between women with RPL and control women (Table 5).

Discussion

Overview of the association of ADIPOQ SNPs with RPL

ADIPOQ is a highly polymorphic gene (<https://www.ncbi.nlm.nih.gov/gene/9370>), and 2376 variants were identified, some of which modulate circulating adiponectin concentrations [18, 23, 25]. We investigated the association between RPL and variants in the intergenic, 5'-near gene, promoter, introns, exons, and 3'-UTR regions of ADIPOQ gene. Among the 14 tested variants, significant association with RPL risk was seen with intergenic (rs4632532), intronic (rs7649121, rs2241767), and exonic (rs2241766, rs1501299) ADIPOQ variants. This is the first report that identified these ADIPOQ variants as RPL at-risk loci, further supporting a role for adiponectin in the pathogenesis of RPL.

Significance of the findings

The association between (exonic) rs1501299 (+276G>T) and RPL seen here was in apparent disagreement with a study on North Indian women, which reported marginal association of the +276G>T variant ($P=0.0360$) with reduced risk of RPL [16]. This discrepancy is likely attributed to differences in ethnic background of study participants, study selection criteria and genotyping conditions. The rs1501299 (+276G>T) variant was associated with

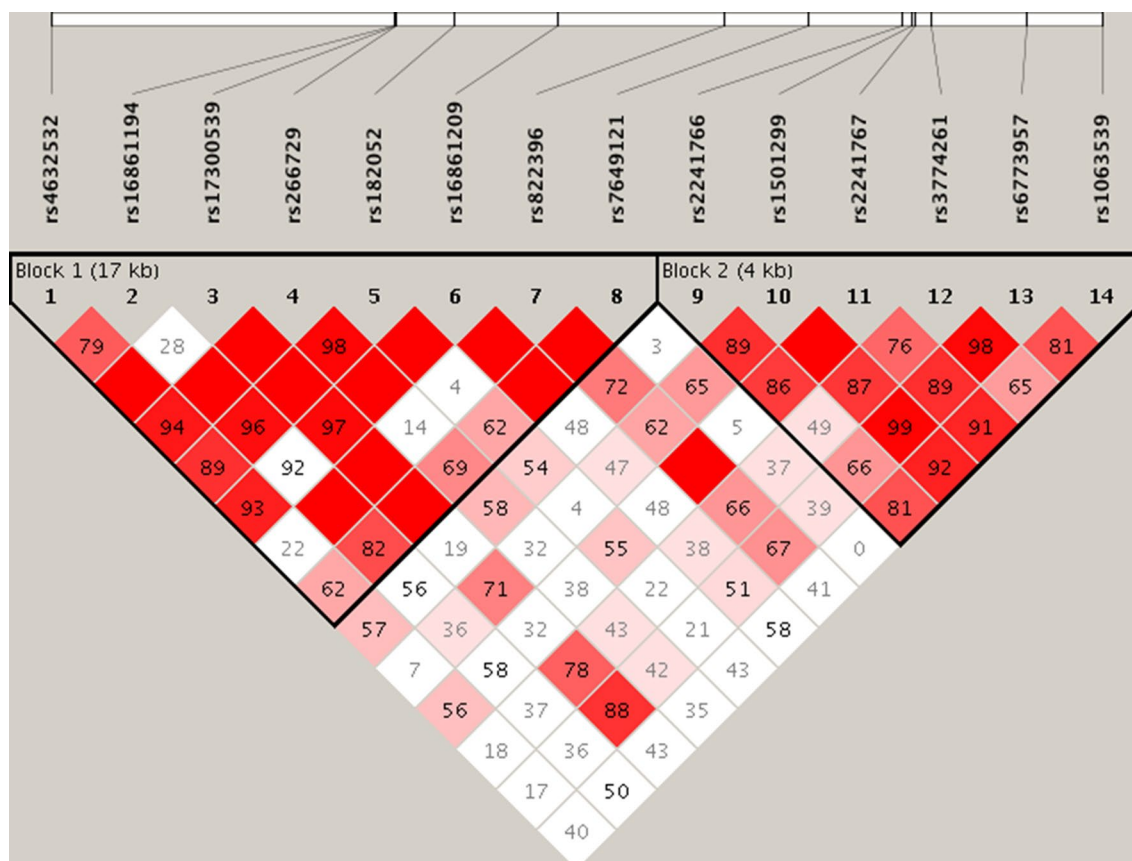


Fig. 1 Haploview plot of *ADIPOQ* SNP analyzed. The relative positions of *ADIPOQ* SNPs (Build 37.3) are displayed, along with the basic gene structure, above the Haploview diagram. The relative LD between pairs of *ADIPOQ* SNPs is color-indicated. This was based on D' , i.e. normalized linkage disequilibrium measure or D divided by the theoretical maximum for the observed allele frequencies, multiplied by 100. Values close to zero indicate no LD, while values approaching 100 indicate full LD. The red colored square represents varying degrees of LD < 1 and LOD (logarithm of odds) > 2 scores; darker shades indicating stronger LD

altered adiponectin serum levels [26], and with hypertensive disorder complicating pregnancy (HDCP) in Chinese subjects [27].

In addition to rs1501299, rs266729 was associated with GDM in Polish [22] and Bulgarian [20], but not Brazilian women [21]; its minor (G) allele conferring protection against GDM [20]. Functionally, rs266729 was linked with altered adiponectin levels in Caucasians [17, 28], which was attributed to the capacity of its minor (G) allele to destroy Sp1 transcription factor DNA binding in the *ADIPOQ* promoter region [29].

ADIPOQ rs7649121 was linked with reduced risk of coronary heart disease in Chinese [30]. In addition, rs17300539 was reportedly associated with reduced adiponectin levels in white women [31], and in T2DM patients [32]. Our study is the first to demonstrate positive association between rs7649121 and rs17300539 and RPL. On the other hand, rs2241767 was negatively associated with RPL in our cohort. This variant was

associated with T2DM in Tunisian Arabs [33], and with insulin resistance in Mexican Americans [34].

ADIPOQ rs2241766 variant is present in exon 2, and its functional attributes are not understood, although it does not alter target amino acid sequence. It was reported that the carriage of rs2241766 is associated with altered mRNA splicing and/or stability [26]. While a number of studies documented association of (exonic) rs2241766 variant with pregnancy complications [21, 35], including PCOS in Brazilian [21], but not Chinese [36] women, it was not associated with RPL in our cohort. This was in agreement with the earlier study on non-obese North Indian women [16], which documented lack of association of rs2241766 with RPL.

Obesity as risk factor of RPL: adiponectin connection

Obesity is a major contributor to infertility, and is associated with reduced success of assisted reproductive

Table 5 Haplotype frequencies across 14 *ADIPOQ* SNPs analyzed^a

Haplotype ^b	Frequency	Case:control frequency	χ^2	<i>P</i>
Block 1 TAGCGCAA	0.433	0.369; 0.499	15.64	7.6×10^{-5}
<u>CAGG</u> ACA <u>I</u>	0.120	0.149; 0.089	7.86	5.1×10^{-3}
TAGCGC <u>G</u> A	0.086	0.082; 0.091	0.19	0.661
<u>C</u> G <u>G</u> C <u>A</u> CAA	0.068	0.076; 0.059	1.01	0.314
T <u>A</u> A <u>C</u> GAAA	0.067	0.087; 0.046	6.27	0.012
<u>C</u> A <u>G</u> <u>G</u> A <u>C</u> G <u>A</u>	0.048	0.053; 0.043	0.52	0.471
TAGCG <u>A</u> AA	0.046	0.054; 0.038	1.23	0.268
<u>C</u> A <u>G</u> <u>G</u> A <u>C</u> AA	0.042	0.051; 0.032	2.14	0.144
TAGCGC <u>A</u> I	0.038	0.037; 0.040	0.04	0.837
Block 2 TCAG <u>A</u> G	0.445	0.475; 0.414	3.71	0.054
T <u>A</u> A <u>A</u> G <u>G</u>	0.291	0.295; 0.287	0.06	0.801
<u>G</u> C <u>G</u> A <u>G</u> C	0.125	0.119; 0.132	0.36	0.546
T <u>C</u> A <u>A</u> G <u>G</u>	0.029	0.023; 0.035	1.40	0.236
<u>G</u> C <u>A</u> G <u>A</u> G	0.021	0.015; 0.026	1.43	0.231

Italicface indicates statistical significance

^a *ADIPOQ* Block 1: rs4632532- rs16861194- rs17300539- rs266729- rs182052- rs16861209- rs822396- rs7649121, Block 2: rs2241766- rs1501299- rs2241767- rs3774261- rs6773957- rs1063539 haplotypes

^b Underlined indicates minor allele

technologies [4, 37]. Given the association of reduced adiponectin to obesity and insulin resistance, a role for adiponectin in promoting successful pregnancy was suggested [14, 15]. There is a solid link between adiposity, obesity, insulin resistance and RPL [38]. Obesity was consistently associated with conditions linked with poor pregnancy outcome, including idiopathic RPL [3, 38].

Adiponectin was proposed to constitute an at-risk modulator of RPL. An earlier systematic review involving 24,738 women, concluded that obesity increases the risk of sporadic miscarriage [37], and was attributed to reduced adiponectin secretion [39, 40]. By controlling steroidogenesis and expression of genes involved in ovulation [13, 41], and induction of metabolic changes linked with adipose tissues dysfunction, particularly in genetically predisposed women [42], a central role of adiponectin in pregnancy outcome was established. In our hands, RPL-susceptible and—protective *ADIPO* SNP and haplotypes were identified, when unselected RPL cases were compared to control women, and also when RPL cases were stratified according to obesity.

Furthermore, we demonstrated that rs16861209, rs182052, and rs7649121 amplify RPL risk in non-obese women. This was in contrast to rs1063539 which was negatively associated with RPL, and presumably

having a protective effect on pregnancy. On the other side, in obese subjects, rs4632532, rs182052, and rs7649121 were linked with increased RPL susceptibility, while rs2241766 was generally protective of RPL. This underscores the need for controlling for modifiable covariates in genetic association studies.

Study strengths and shortcomings

The strength of our study is that it is sufficiently powered, and that RPL cases and control women were matched according to ethnicity, which reduces the problems of ethnic differences inherent in genetic association studies. Another strength is controlling for potential covariates. Our study has also some shortcomings. We could not measure serum adiponectin levels in cases and control women, which did not allow for addressing the functionality of this association, thus could not ascertain genotype–phenotype correlations. Another limitation lies in its retrospective nature, which prompts speculation on cause-effect relationship.

Conclusion

We demonstrated association of *ADIPOQ* rs4632532, and rs2241766 with RPL in obese women, and rs1063539 and rs16861209 in non-obese women, and both rs182052 and rs7649121 independent of BMI changes. Future studies on additional *ADIPOQ* variants, and populations of related and distant ethnic origin are needed to support, or rule out association of *ADIPOQ* variants with altered adiponectin secretion and risk of RPL.

Abbreviations

ADIPOQ: adiponectin gene; BP: blood pressure; CI: confidence intervals; GDM: gestational diabetes mellitus; HWE: Hardy–Weinberg equilibrium; LD: linkage disequilibrium; MAF: minor allele frequency; OR: odds ratios; PCOS: polycystic ovarian syndrome; PE: preeclampsia; RPL: recurrent pregnancy loss; SNP: single nucleotide polymorphisms.

Authors' contributions

MD, WB: Sample processing, drafting of manuscript, RRF: Patient screening and referral, MM: Genotyping assays, WYA: Project leader, finalizing the manuscript. All authors read and approved the final manuscript.

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Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Consent for publication

Not applicable.

Ethics approval and consent to participate

The study protocol was approved by Arabian Gulf University Research and Ethics Committee (IRB approval: 35-PI-01/15, granted on 17 October 2014), and was done according to Helsinki II Declaration. All patients provided informed written consent before blood sampling.

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