

REVIEW

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Mechanisms involved in breast cancer liver metastasis

Rui Ma¹, Yili Feng², Shuang Lin², Jiang Chen², Hui Lin², Xiao Liang², Heming Zheng² and Xiujun Cai^{2*}

Abstract

Liver metastasis is a frequent occurrence in patients with breast cancer; however, the available treatments are limited and ineffective. While liver-specific homing of breast cancer cells is an important feature of metastasis, the formation of liver metastases is not random. Indeed, breast cancer cell factors contribute to the liver microenvironment. Major breakthroughs have been achieved recently in understanding breast cancer liver metastasis (BCLM). The process of liver metastasis consists of multiple steps and involves various factors from breast cancer cells and the liver microenvironment. A further understanding of the roles of breast cancer cells and the liver microenvironment is crucial to guide future work in clinical treatments. In this review we discuss the contribution of breast cancer cells and the liver microenvironment to liver metastasis, with the aim to improve therapeutic efficacy for patients with BCLM.

Keywords: Breast cancer, Liver metastasis, Microenvironment

Introduction

Breast cancer is the leading cause of cancer-related deaths in female patients worldwide [1]. Breast cancer has been divided into at least five subtypes, as follows: luminal A, luminal B, human epithelial growth receptor type 2 (HER-2), basal-like, and claudin-low [2]. It has been reported that the 5-year survival rate for primary breast cancer is 99%. Nevertheless, approximately one-third of breast cancer patients will present with distant non-nodal metastases, and the 5-year survival rate decreases to 23% once distant metastases have developed [3]. Breast cancer mainly metastasizes to the bony skeleton, lungs, liver, and brain via the circulation; the liver is a common metastatic site for solid cancers and represents the third most common site for breast cancer [4]. If breast cancer liver metastasis (BCLM) is left untreated, the survival time is only 4–8 months [5]. Current treatments for BCLM are based on a strategy of systemic chemotherapy, endocrine- or HER2-targeted therapy (depending on estrogen receptor [ER], progesterone receptor [PR], and HER-2 status), and palliative therapy, such as radiation [6,7]. Nevertheless, it is important to note that some patients exhibit *de novo* resistance or eventually become

resistant to endocrine therapy [8]. Moreover, a poor response to chemotherapy accounts for much of the high mortality in patients with BCLM [9].

The formation and growth of breast cancer cells in the liver is a complex process. The most widely accepted model for metastasis is the “seed and soil” hypothesis, which was postulated by Stephen Paget in 1889 [10]. Paget speculated that organ metastases form merely when the seed (disseminated tumor cells) and soil (secondary organ) are compatible [10,11]. Semenza separated the process of blood vessel metastasis of breast cancer into the following steps: intravasation, circulation, margination, extravasation, and colonization [12]. The molecular mechanisms underlying breast cancer metastasis have been reported for breast cancer dissemination to the lungs and bone [13–20]; however, the molecular mechanisms for liver metastasis have not been completely described [21]. Moreover, the hepatic microenvironment and liver sinusoidal structure is crucial for the initial arrest of breast cancer and progression within the liver. Thus, exploring the mechanisms underlying liver metastasis in breast cancer patients is essential for developing more effective therapies. A further understanding of the roles of breast cancer cells and the liver microenvironment in early breast cancer metastasis is crucial for the development of effective BCLM therapies [10,22]. Therefore, this review will discuss the molecular

* Correspondence: cxjzu@hotmail.com

²Department of General Surgery, Institute of Minimally Invasive Surgery, Sir Run Run Shaw Hospital, College of Medicine, Zhejiang University, Hangzhou, Zhejiang 310016, China

Full list of author information is available at the end of the article

mechanisms underlying liver metastasis of breast cancer to guide future work in clinical treatments.

Factors associated with breast cancer cell metastasis to the liver

Inflammatory factors

Several studies have shown that the inflammatory response correlates with the liver metastatic potential of some tumors [23,24]. The inflammatory factor, TNF- α , can trigger the expression of E-selectin in endothelial cells, including liver sinusoidal endothelium cells [25,26]. It has been reported that breast cancer cells have the ability to initiate an inflammatory cascade, which increases adhesiveness to liver sinusoidal endothelium cells, similar to that induced by colorectal and lung cancers [27]. Although the process of tumor cell attachment to the endothelium during metastasis is multifactorial, the production of TNF- α -induced endothelial E-selectin in tumor cells appears to be a key step in the BCLM process [28]. Asgeirsson et al. [29] reported that induction of IL-6 decreased cell adhesion in three breast cancer cell lines, and was associated with reduced E-cadherin expression. Moreover, patients with breast cancer liver metastases had significantly higher IL-6 levels. Therefore, it appears that breast cancer cells create a pro-inflammatory microenvironment which triggers adhesion and invasion of tumor cells into the liver by secreting a number of cytokines.

Chemokines and chemokine receptors

Breast cancer cells express several chemokine receptors that initiate liver metastasis, of which C-X-C Chemokine Receptor type 4 (CXCR4) is the most common. Moreover, the ligand of CXCR4, stromal-derived factor 1- α (SDF-1, CXCL12), is also highly expressed in the liver [30], indicating that the CXCL12/CXCR4 interaction might contribute to BCLM. It has also been reported that CXCR4 plays an important role in modulating breast-to-liver metastasis through integrin-adhesion-receptor signaling [31]. The availability of chemokine receptors on tumor cell surfaces, the presence of specific ligands within the microenvironment of potential target organs, and the suitability of the extracellular matrix (ECM) composition appear to be required for successful extravasation of breast cancer cells in the early metastatic process [32]. Furthermore, expression of CXCR4 increases the risk of metastasis to the liver in patients with axillary node-positive primary breast cancer [33]. In tumor-bearing mice, CC chemokine ligand 2 (CCL2) neutralizing antibodies inhibit the growth and liver metastases of primary breast cancer by reducing cell proliferation, survival, and tumor-associated macrophage (TAM) recruitment. These results have also revealed that CCL2 can enhance primary breast cancer liver metastasis in a TAM-dependent manner [34]. Stormes et al. [35] strongly support the notion that

inhibition of tumor-derived CCL5 can inhibit the capability of liver metastasis in breast cancer cells. Similar analyses have shown that the release of CCL5 by cells of the tumor microenvironment promotes the metastatic spread to the liver of breast cancer cells [36]. Moreover, Mi et al. [37] have demonstrated that induced mesenchymal stromal cells (MSCs) produce CCL5, and significantly promote breast cancer cell migration to the liver *in vivo* and *in vitro*. Therefore, some chemokine receptors can enhance breast cancer metastasis to the liver. Porter et al. [38] reported, however, that many chemokines are lost in breast cancer, including CXCL1, CXCL2, CXCL5, CXCL6, CXCL8, CXCL20, CX3CL1, CCL2, and CCL7. Whether or not some of these chemokines are involved in BCLM warrants further study. Taken together, chemokines and their receptors are the regulators involved in the process of BCLM.

Cell adhesion molecules

Cadherins

E-cadherin expression in liver metastatic sites is due to loss of promoter methylation. Breast cancer cells that re-express E-cadherin revert back to an epithelial phenotype [39-42]. In primary cancer, the epithelial-to-mesenchymal transition (EMT) of cancer cells contributes to increased invasion and dissemination [43]. Once cancer cells have seeded the metastatic site, a mesenchymal-to-epithelial transition (MET) occurs, leading to the colonization and growth of metastatic foci [44]. Expression of the cell adhesion molecule, E-cadherin, in breast cancer cells can facilitate breast cancer cell adhesion to hepatocytes for seeding in the liver [9,43]. Moreover, some results have shown that breast cancer cells that express E-cadherin are able to form liver metastases, while E-cadherin-negative cancer cells merely form primary tumors [45,46]. Intriguingly, Chao et al. [9] showed that the liver microenvironment can induce breast cancer cells to re-express E-cadherin and cause MET. This phenotypic change has the potential to alter cell behavior, and thus may be a critical step for cells to survive at metastatic sites within the liver. Take together, these results suggest that re-expression of E-cadherin, accompanied by a partial MET in the liver, increases post-extravasation survival of metastatic cancer cells and may help to elucidate why chemotherapy commonly fails to treat BCLM.

Previous studies have shown that N-cadherin is up-regulated in more invasive and less differentiated breast cancer cell lines that lack E-cadherin expression [47,48]. The study by Hazan et al. [49] showed that breast cancer expressing N-cadherin grew slower, on average, and the two clones with the highest levels of N-cadherin formed liver metastases in almost all mice injected with breast cancer cells. In addition, N-cadherin activates a metastatic signaling pathway coordinated by fibroblast growth factor receptor (FGFR) and matrix metalloproteinase-9 (MMP-9)

to overcome the suppressive effects regulated by E-cadherin. Thus, these studies provide a basis for N-cadherin-mediated liver metastasis of breast cancer, whereas E-cadherin suppresses liver metastases, and offer new insights into the diagnostic or therapeutic applications for BCLM.

Integrin complexes

Heterodimeric transmembrane receptors of the integrins family are important components of the ECM. Eighteen α -subunits and 8 β -subunits have been characterized and form 24 different integrins [50], including laminin (LN), collagen (Col), fibrinogen, and vitronectin (VN) [51]. Each integrin heterodimer binds to a specific ligand within the ECM. By using bidirectional “outside-in” and “inside-out” signaling, integrin complexes regulate multiple biological processes, such as adhesion, apoptosis, proliferation, differentiation, migration, invasion, and metastasis [52]. It has been reported that use of an antagonist of $\alpha 2\beta 1$ complexes can reduce the extravasation of colorectal and hepatocellular carcinoma cell lines into the liver and micrometastatic sites [53]. Moreover, the functional integrin complexes (fibronectin and collagen IV receptors) are recruited, assembled, and thus increased on the cell surface in liver metastatic breast cancer cells. Neutralizing antibody targeting $\alpha 5\beta 1$ or $\alpha 2\beta 1$ complexes can block claudin-2-mediated adhesion to fibronectin and type IV collagen, and reduce the ability of breast cancer cells to metastasize to the liver [54]. Therefore, $\alpha 2\beta 1$ or $\alpha 5\beta 1$ complexes can promote the ability of breast cancer cells to metastasize to the liver, at least partially, via the claudin-2 signaling pathway.

Ig-SF

Epithelial cell adhesion molecule (EpCAM/CD326), also a transmembrane protein, plays a variety of roles in cell proliferation, adhesion, migration, and tissue maintenance [55,56]. EpCAM is expressed at low levels in luminal epithelial cells of benign breast tissues. Nevertheless, EpCAM is overexpressed in many carcinomas, including breast cancer; a recent study suggested that p53 dysfunction may serve to explain this phenomenon [57]. In patients with node-positive primary breast cancer, elevated EpCAM expression correlates with diminished overall survival [58,59], suggesting that overexpression of EpCAM promotes cancer progression and metastasis. Litvinov et al. [60,61] also found that EpCAM can act as an antagonist impairing E-cadherin function via disruption of the α -catenin/F-actin link, and loosening tight cell-cell adhesions. In addition, EpCAM is highly expressed in MBCs, including the sites of liver metastases, compared to unmatched, surgically resected primary breast cancers [61]. Therefore, EpCAM might serve as a promising therapeutic target for BCLM.

CD44

It has been reported that cancer stem cells (CSCs) within tumors have cancer-initiating potential and metastatic capability [62]. High levels of CD44 expressed by CSCs are believed to be involved in adhesion, invasion, apoptosis resistance, and metastasis [63-65]. Moreover, breast cancer cells expressing high levels of CD44 and low levels of CD24 maintain stemness properties [62,66]. Erin et al. [67] found that CD44 expression was highest in cells that metastasized to the liver, and liver tropism of breast cancer is driven by CSCs. In addition, it has been demonstrated that serum CD44 v5 and v6 are released by breast cancer cells and increased CD44 v6 serum levels are preferentially detected in patients with liver metastases [68,69]. Furthermore, Ouhitt et al. [70] showed that CD44 promotes breast tumor invasion and metastasis to the liver. Together, high levels of CD44 expressed by CSCs and breast cancer cells are significantly correlated with BCLM.

Based on the above results, it appears that cell adhesion molecules play important roles in BCLM (Table 1); however, studies involving cell adhesion molecules that effect BCLM is still limited, and uncovering the mechanisms would provide novel therapeutic approaches in drug design and cancer therapy.

Role of claudins

In normal epithelia, claudins are key transmembrane proteins within the tight junction complex that participate in homo- and hetero-typic interactions between adjacent cells [71]. The roles of claudin proteins in breast cancer progression are complicated, and claudin-2 expression is down-regulated compared with normal tissues. Moreover, decreased levels of claudin-2 are observed in high-grade breast cancer, suggesting that claudin-2 plays a suppressive role in breast cancer [72]. Moreover, Kimbung et al. [73] reported that claudin-2 is a prognostic biomarker that not only predicts the likelihood of a breast cancer recurrence, but more interestingly, the metastatic potential of breast cancer to the liver. Furthermore, Tabariès et al. [54,74] demonstrated that elevated levels of claudin-2 expression, or selection for pre-existing claudin-2-positive breast cancer cells within liver metastases may serve to enhance the survival of breast cancer cells by promoting interactions between the tumor cell and resident hepatocytes. Claudin-2 mediates breast cancer metastasis to the liver, at least partially, by enhancing adhesion to ECM proteins, such as fibronectin and type IV collagen, which are abundant in the liver. Immunohistochemical analyses have shown that claudin-2 is detected in all liver metastases and weakly expressed in primary human breast cancers. Take together, these results revealed novel roles of claudin-2 in promoting breast cancer adhesion to the ECM and breast cancer metastasis to the liver.

Table 1 Cell adhesion molecules (CAMs) involved in breast cancer liver metastasis

CAM family	Expression in primary cancer	Role	Expression in BCLM	Role
Cadherins				
E-cadherin	↓Breast cancer cells	Adhesion, invasion, EMT	↑Breast cancer cells [9,43,45,46]	MET
N-cadherin	↑Breast cancer cells, ↑Endothelium cells	Angiogenesis, EMT	↑Breast cancer cells [49]	MET
Integrin complexes				
α2β1	↓Breast cancer cells, ↑Endothelium cells	Adhesion, migration, invasion, angiogenesis	↑Breast cancer cells [54]	liver metastasis, tumor angiogenesis
α5β1	↑Breast cancer cells, ↑Endothelium cells	Adhesion, migration, invasion	↑Breast cancer cells [54]	liver metastasis, tumor angiogenesis
Ig-SF				
EpCAM	↑Breast cancer cells	Migration, invasion	↑Breast cancer cells [61]	liver metastasis
CD44				
	↓Breast cancer cells (CD44s); ↑Breast cancer cells (CD44v3, CD44v4, CD44v5, CD44v6)	adhesion, invasion	↑Breast cancer cells [67,68]	Liver metastasis

Note: ↑, up-regulation; ↓, down-regulation; EMT, epithelial to mesenchymal transition; MET, mesenchymal to epithelial reverting transition.

The up-regulation of claudin-3 and -4 is correlated with poor prognosis and the breast cancer basal-like subtype [75-77]. A recent study demonstrated that the loss of claudin-4 and -7 promoted liver metastasis of breast cancer cells in Balb-c mice [78]. We speculate that the discrepant roles of claudin-4 with poor prognosis in different studies might be due to different subtypes or species. Intriguingly, a 'claudin-low' subtype of breast cancer has recently been identified in primary human breast cancers. This subtype possesses features similar to, but distinct from the breast cancer basal subtype defined previously [79]; the breast cancer claudin-low and basal subtypes are functionally coupled by EMT characteristics, enhanced cancer stem cell-like features, and resistance to chemotherapy [80-84]. Nevertheless, the patterns of expression of claudins in the claudin-low subtype of breast cancer have not been described.

In summary, the above findings imply that claudin-2, -4, and -7 are all necessary and sufficient for the ability of breast cancer cells to colonize and grow in the liver. It is important to delineate the mechanism underlying preferential metastasis of breast cancer to the liver.

Breast cancer subtypes

It has been reported that five major subtypes of breast cancer have different abilities to metastasize to distant organs, and share pathways with the preferred metastatic sites. Patients with bone relapses have the luminal subtypes of breast cancer most frequently. The HER-2 subtype may metastasize to bone via processes that differ from the luminal subtypes. Moreover, the basal subtype often metastasizes to the brain and lungs. However, it fails to reach statistical significance in patients with liver relapse [85]. Rodriguez-Pinilla et al. [86] reported

that the basal-like subtype metastasizes more frequently to the lungs and other visceral organs, such as the brain and liver, but no bone metastases were detected. Moreover, recent studies [87-89] in patients from eastern Europe, Asia, and the US have suggested a high incidence of brain metastases arising from basal-like tumors (ER-/PR-/HER2-, and usually identified as "triple-negative" breast cancer [TNBC]) [90], but it has been reported that p53-negative TNBC has an increased tendency to develop lung metastases [91]. Furthermore, Duan et al. [92] showed that the breast cancer subtype is an independent prognostic predictor for patients with breast cancer metastases to the liver. Survival after liver metastases arising from TNBC is 21 months compared to 30, 32, and 41 months for patients with the HER-2, luminal B, and luminal A subtypes; liver metastases from TNBC has the worst prognosis. Therefore, novel agents controlling liver metastases in patients with TNBC are needed. Taken together, different subtypes of breast cancer have preferred metastatic organs; however, no conclusive evidence exists to mechanistically link any specific subtype to BCLM development.

Factors associated with the liver microenvironment

Hypoxia-inducible factor-regulated genes

Hypoxia-inducible factors (HIFs) activate the transcription of target genes that are involved in many aspects of breast cancer progression, such as angiogenesis, metabolic reprogramming, local tissue invasion, and metastasis [12,93]. It has been reported that HIFs not only activate lysyl oxidase (LOX) directly to inhibit liver metastases, but osteopontin (OPN), vascular endothelial growth factor (VEGF), and TWIST promote BCLM (Table 2). It has been suggested that some of the

Table 2 Hypoxia-inducible factor (HIF)-regulated genes in breast cancer liver metastasis

Gene	HIF regulation	Role in BCLM
LOX [94]	HIF-1 and HIF-2	Inhibition
OPN [37,69]	HIF-1 (HIF-2 not tested)	Promotion
Twist [95,116]	HIF-1 (HIF-2 not tested)	Promotion
VEGF [95]	HIF-1 and HIF-2	Promotion

hypoxia-inducible factor-regulated genes contribute to BCLM [37,69,94,95].

LOX

LOX is an amine oxidase that contributes to the formation of the ECM. LOX is secreted by fibrinogenic cells and residues in collagen and elastin to maintain the structural stabilization of the ECM [96]. Of note, LOX expression has paradoxical roles in tumor suppression and tumor progression, depending on cellular location, type, and transformation status [96-101]. Erler et al. [94] proposed that hypoxia-induced LOX has a key function in the metastasis of breast cancer cells. Although no marked inhibition effects of LOX on primary breast cancer growth was observed, Erler et al. [94] found significant effects on growth in metastatic sites within the liver. These data indicate that the effects of LOX on cell adhesion, migration, invasion, and three-dimensional growth are more crucial for liver metastatic growth than primary breast cancers. This study may provide mechanistic evidence for hypoxia-driven liver metastases and support the therapeutic target of LOX in the prevention and treatment of BCLM.

OPN

HIF-induced OPN is a secreted phosphoprotein functioning as a cell attachment protein by binding two cell adhesion molecules ($\alpha\text{v}\beta\text{3}$ integrin and CD44) [102-104]. It has been reported that OPN is overexpressed in tumors and elevated serum OPN levels are associated with advanced metastatic cancer [105-108]. Gain- and loss-of-function assays have demonstrated the critical role for OPN in tumor metastases of colon, liver, and breast cancers [69]. Mi et al. [37] revealed that MSCs extracted from metastatic sites exhibit significantly increased expression of cancer-associated fibroblast (CAF) markers, such as α -smooth muscle actin (α -SMA), tenascin-c, CXCL12, and fibroblast-specific protein (FSP)-1. OPN expression promotes tumor growth and metastasis by activating the expression of CCL5, MMPs, and CAF in MSCs. Thus, the transformation of MSCs to CAF can be mediated by OPN in the tumor microenvironment. In addition, the current study showed that the expression of CAF markers was also significantly increased in the liver metastases sites. These findings suggest a novel mechanism

by which OPN affects BCLM by transforming MSCs into a CAF.

VEGF and TWIST

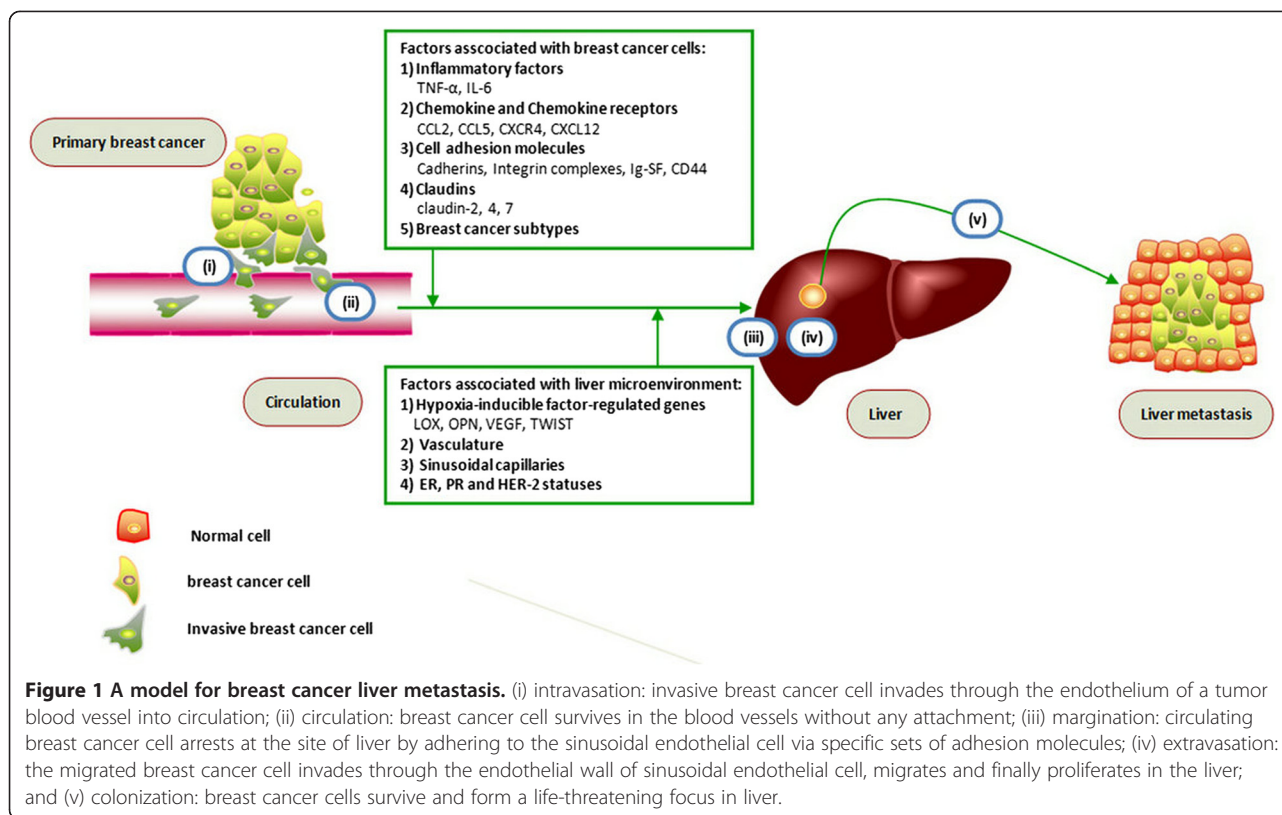
Cancer cells express a number of angiogenic factors. VEGFs are the key mediator of neovascularization in tumors [109]. Inhibition of VEGF-A/VEGFR-2 and VEGF-C/VEGFR-3 signals has been shown to suppress breast cancer progression and lung metastases [110]. Chien et al. [111] found that inhibition of VEGFR/FGFR kinases drastically reduce the formation of liver metastases and decreased primary breast cancer growth. TWIST is a basic helix-loop-helix transcription factor. TWIST mainly regulates gastrulation and mesoderm specification [112,113]. Recently, TWIST has been shown to play an important role in mediating cancer metastasis [114,115]. TWIST is a downstream target of HIF-1 and has an important role in metastatic phenotypes induced by hypoxia or overexpression of HIF-1 α in breast cancer cell lines (MCF-7). HIF-1 α promotes hypoxia-induced breast cancer progression and metastasis through the direct activation of TWIST expression [116].

In a previous report, inhibition of HIF-1 α by 2-benzoyl-3-phenyl-6, 7-dichloroquinoxaline 1, 4-dioxide (DCQ) was shown to block VEGF secretion and invasion in MCF-7 and led to the inhibition of TWIST expression in MDA-MB-231. DCQ exhibits robust anti-tumor activity in MDA-MB-231 breast cancer mouse xenografts. Also, DCQ reduces metastatic dissemination to the liver, leading to prolonged animal survival [95]. Therefore, HIF-1 α can promote BCLM through VEGF and TWIST signaling pathways.

Role of the vasculature might be essential for liver metastasis

Vermeulen et al. [117] reported that colorectal cancer metastases to the liver grow according to three different histologic patterns, termed 'pushing', 'replacement', and 'desmoplastic' growth patterns. The results of subsequent studies [118,119] involving liver metastases of colorectal and breast cancers showed that the different grow patterns have different angiogenic properties. The replacement pattern grows by co-opting the stroma, without induction of hypoxia or angiogenesis and thus minimal perturbation of the liver architecture; however, the pushing and desmoplastic patterns grow in an angiogenic fashion, which is at least in part hypoxia-driven. Furthermore, the pattern of replacement growth in a non-angiogenic process is even more prevalent in breast cancer than colorectal cancer, and the process induces neither hypoxia nor vascular leakage.

Martin et al. [120] has shown that the majority of early metastatic foci in the liver contain few cells, even 12 days after breast cancer cell injection. Only a few foci were able to develop into micrometastatic lesions with a patent



vasculature, thus suggesting that lesions that utilize an existent patent blood supply can thrive in the liver microenvironment, while the remaining foci without a vascular supply remain dormant in the liver. Naumov et al. [121] suggested that tumors are dependent on angiogenesis for progressive growth and remain harmless to the organism at the non-angiogenic dormant stage. Furthermore, the expansion of tumor mass is associated with recruiting endothelial cells after the cancer tissues undergo a switch from a non-angiogenic dormant phenotype to the angiogenic phenotype [122].

Therefore, we speculate that breast cancer metastases to the liver grow mainly by the replacement pattern with a non-angiogenic process in the initial stage, and the vasculature is crucial for thriving in the sites of liver metastases in the late stage.

Role of sinusoidal capillaries

Previous studies have reported that the initial arrest of cancer cells in the sinusoids of the liver is restricted by the sizes of cancer cells [123,124]. Haier et al. [125] has determined that tumor cells adhere to sinusoidal capillaries, the internal diameter of which is larger than the tumor cells. Unique structural features of liver, including the existence of a fenestrated endothelium (sinusoidal endothelium) and lack of an organized sub-endothelial basement membrane, have a great impact on the interactions between

breast cancer cells and the liver microenvironment. Of great interest, the fenestrated endothelium controls liver-specific microvascular exchange and impacts the ability of cells to transmigrate through the vessels into the liver [126]. Moreover, previous studies have revealed that breast cancer cells extend cellular projections through the fenestrated endothelium into the space of Disse on seeding the liver, which makes direct contact with hepatocytes [127]. In addition, Martin et al. [120] has found that breast cancer cells are bound to vessels with clear vascular labeling in the sites of liver metastases. Thus, sinusoidal capillaries play a significant role in the initial arrest of breast cancer.

Changes in hormonal receptor status and HER-2

ER, PR, and HER-2 status is essential in determining the use and evaluating the effect of adjuvant hormone therapy, molecular targeted therapy, and even chemotherapy. Koo et al. [128] conducted a study to assess the status of ER, PR, and HER-2 in primary and metastatic breast cancers and determined the relationship between ER, PR, and HER-2 and organ-specific metastases of breast cancer. The data showed that ER+ or PR+/HER-2-(luminal A) subtypes were predominant in the sites of liver metastases (75.0%). Increased phosphorylation of HER-2 appears to be extremely important for the establishment of breast cancer liver metastases [129]. Moreover,

a high serum HER2 level or lack of ERs independently doubles the relative risk of progression and mortality [130].

Nevertheless, the ER, PR, and HER-2 status between primary breast cancers and liver metastatic foci can be changed after treatment, but are stable in most cases during the liver metastatic process [131]. Botteri et al. [132] conducted a retrospective study of patients with BCLM and found a positive relationship between liver biopsy findings and survival in patients with early metastases. Moreover, another study showed that biopsies of metastases are useful for the reassessment of the metastatic sites to define a more effective treatment strategy for patients with BCLM [133]. Thus, the ER, PR, and HER-2 status needs to be reassessed by biopsy when liver metastases occur.

A model for breast cancer liver metastasis

An increasingly sophisticated understanding of breast cancer liver metastasis is emerging. It appears that BCLM is mainly associated with specific subtypes in patients with breast cancer [85,86,92]; however, no direct correlation between subtypes and BCLM has been found. By combining the knowledge from the extant research, we propose a model for breast cancer liver metastasis (Figure 1), as follows: (i) intravasation: invasive breast cancer cells invade via the endothelium of a tumor blood vessel into the circulation; (ii) circulation: breast cancer cells survive in the blood vessels and lack of cell-cell or cell-matrix attachments; (iii) margination: CTCs arrest at the liver site by adhering to the sinusoidal endothelial cell via specific sets of adhesion molecules, such as cadherins, integrins, Ig-SF, and CD44; (iv) extravasation: the migrated breast cancer cells invade through the endothelial wall of sinusoidal endothelial cells, migrates, and finally proliferates in the liver (in this process, the diameter of the sinusoidal endothelium and the lack of an organized sub-endothelial basement membrane have a great impact on breast cancer cell migration); and (v) colonization: breast cancer cells survive and form a life-threatening macrometastatic focus in the liver microenvironment by mediating hypoxia-inducible factor-regulated genes (LOX, OPN, VEGF, and TWIST), the status of ER, PR, and HER-2 expression, and angiogenesis for breast cancer cells.

Conclusions

The process of BCLM includes several steps and is influenced by various factors. Although major breakthroughs have been achieved recently in understanding of BCLM, no effective therapies have been developed. Further understanding of the roles of breast cancer cells and the liver microenvironment will open a new window to guide future work in clinical treatments.

Abbreviations

BCLM: Breast cancer liver metastasis; CAF: cancer-associated fibroblast; CCL2: CC chemokine ligand 2; Col: Collagen; CSCs: Cancer stem cells; CXCR4: C-X-C Chemokine Receptor type 4; DCQ: 2-benzoyl-3-phenyl-6,7-dichloroquinoxaline 1, 4-dioxide; ECM: Extracellular matrix; EMT: Epithelial to mesenchymal transition; EpCAM: Epithelial cell adhesion molecule; ER: Estrogen receptor; FGFR: Fibroblast growth factor receptor; FSP: Fibroblast-specific protein; HER2: Human epidermal growth factor receptor-2; HIF: Hypoxia-inducible factor; LN: Laminin; LOX: Lysyl oxidase; MET: Mesenchymal to epithelial transition; MMP-9: Matrix metalloproteinases 9; MSC: Mesenchymal stromal cell; OPN: Osteopontin; PR: Progesterone receptor; TAM: Tumor-associated macrophage; TNBC: "triple-negative" breast cancer; VEGF: Vascular endothelial growth factor; VN: Vitronectin; α -SMA: α -smooth muscle actin.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

RM, SL and JC contributed to collection of information, analysis and interpretation of data and writing of the manuscript. YLF and HL contributed to revision of the manuscript. HMZ, XL and XJC contributed to design and revision of the manuscript. All authors read and approved the final manuscript.

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Author details

¹Department of Surgery, Zhejiang University Hospital, Zhejiang University, Hangzhou, Zhejiang 310027, China. ²Department of General Surgery, Institute of Minimally Invasive Surgery, Sir Run Run Shaw Hospital, College of Medicine, Zhejiang University, Hangzhou, Zhejiang 310016, China.

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